

## POSITION PAPER



# Nomenclature of allergic diseases and hypersensitivity reactions: Adapted to modern needs: An EAACI position paper

Marek Jutel<sup>1,2</sup>  | Ioana Agache<sup>3</sup>  | Magdalena Zemelka-Wiacek<sup>1</sup>  | Mübeccel Akdis<sup>4</sup>  |  
 Tomás Chivato<sup>5</sup>  | Stefano del Giacco<sup>6,7</sup>  | Pawel Gajdanowicz<sup>1</sup>  |  
 Ibon Eguiluz Gracia<sup>8</sup>  | Ludger Klimek<sup>9,10</sup>  | Antti Lauerma<sup>11</sup>  | Markus Ollert<sup>12,13</sup>  |  
 Liam O'Mahony<sup>14</sup>  | Jürgen Schwarze<sup>15</sup>  | Mohamed H. Shamji<sup>16,17</sup>  |  
 Isabel Skypala<sup>18,19</sup>  | Oscar Palomares<sup>20</sup>  | Oliver Pfaar<sup>21</sup>  | Maria Jose Torres<sup>8</sup>  |  
 Jonathan A. Bernstein<sup>22</sup>  | Alvaro A. Cruz<sup>23</sup>  | Stephen R. Durham<sup>24</sup>  |  
 Stephen J. Galli<sup>25</sup>  | R. Maximiliano Gómez<sup>26</sup>  | Emma Guttman-Yassky<sup>27</sup>  |  
 Tari Haahtela<sup>28</sup>  | Stephen T. Holgate<sup>29</sup>  | Kenji Izuhara<sup>30</sup>  | Kenji Kabashima<sup>31</sup>  |  
 Désirée E. Larenas-Linnemann<sup>32</sup>  | Erica von Mutius<sup>33,34,35</sup>  | Kari C. Nadeau<sup>36</sup>  |  
 Ruby Pawankar<sup>37</sup>  | Tomas A. E. Platts-Mills<sup>38</sup>  | Scott H. Sicherer<sup>39</sup>  |  
 Hae-Sim Park<sup>40</sup>  | Stefan Vieths<sup>41</sup>  | Gary Wong<sup>42</sup>  | Luo Zhang<sup>43,44</sup>  |  
 M. Beatrice Bilò<sup>45</sup>  | Cezmi A. Akdis<sup>4</sup> 

**Correspondence**

Marek Jutel, Department of Clinical Immunology, Wrocław Medical University, Wrocław, Poland.

Email: [marek.jutel@all-med.wroclaw.pl](mailto:marek.jutel@all-med.wroclaw.pl)

Ioana Agache, Faculty of Medicine, Transylvania University, Brasov, Romania.  
 Email: [ibrumaru@unitbv.ro](mailto:ibrumaru@unitbv.ro)

Cezmi A. Akdis, Swiss Institute of Allergy and Asthma Research (SIAF), University of Zurich, Davos, Switzerland.  
 Email: [cezmi.akdis@siaf.uzh.ch](mailto:cezmi.akdis@siaf.uzh.ch)

**Abstract**

The exponential growth of precision diagnostic tools, including omic technologies, molecular diagnostics, sophisticated genetic and epigenetic editing, imaging and nanotechnologies and patient access to extensive health care, has resulted in vast amounts of unbiased data enabling in-depth disease characterization. New disease endotypes have been identified for various allergic diseases and triggered the gradual transition from a disease description focused on symptoms to identifying biomarkers and intricate pathogenetic and metabolic pathways. Consequently, the current disease taxonomy has to be revised for better categorization. This European Academy of Allergy and Clinical Immunology Position Paper responds to this challenge and provides a modern nomenclature for allergic diseases, which respects the earlier classifications back to the

**Abbreviations:** ACD, allergic contact dermatitis; AD, atopic dermatitis; ADCC, antibody-dependent cellular cytotoxicity; AERD, aspirin-exacerbated respiratory diseases; AGEP, acute generalized exanthematous pustulosis; APC, antigen-presenting cell; AR, allergic rhinitis; ARC, allergic rhinoconjunctivitis; B, B lymphocytes; BAS, basophil; CRS, chronic rhinosinusitis; DRESS, severe drug reaction with eosinophilia and systemic symptoms; EoE, eosinophilic oesophagitis; EOS, eosinophil; FPIES, food protein-induced enterocolitis syndrome; HDM, house dust mite; IFN- $\gamma$ , interferon-gamma; IgE/G/M/A, immunoglobulin type E/G/M/A; IL, interleukin; ILC1/2/3, innate lymphoid cells type 1/2/3; MC, mast cell; MO, monocyte; M $\phi$ , macrophage; NEU, neutrophils; NK, natural killer cell; NK-T, natural killer T cell; NSAID, nonsteroidal anti-inflammatory drugs; sIgE, allergen-specific IgE; SJS, Stevens-Johnson syndrome; T1/T2/T3, type 1/2/3 immune response; Tc1/2/17, T cytotoxic lymphocyte type 1/2/17; TEN, toxic epidermal necrolysis; Tfh, T follicular helper cells; Tfr, follicular regulatory T cells; TGF- $\beta$ , tumour growth factor-beta; Th1/2/9/17/22, T helper lymphocyte type 1/2/9/17/22; TLSP, thymic stromal lymphopoietin; TNF- $\alpha$ , tumour necrosis factor-alpha; TSLP, thymic stromal lymphopoietin.

Marek Jutel and Ioana Agache—joint first co-authorship.

M. Beatrice Bilò and Cezmi A. Akdis—joint senior co-authorship.

© 2023 European Academy of Allergy and Clinical Immunology and John Wiley & Sons Ltd.

early 20th century. Hypersensitivity reactions originally described by Gell and Coombs have been extended into nine different types comprising antibody- (I-III), cell-mediated (IVa-c), tissue-driven mechanisms (V-VI) and direct response to chemicals (VII). Types I-III are linked to classical and newly described clinical conditions. Type IVa-c are specified and detailed according to the current understanding of T1, T2 and T3 responses. Types V-VI involve epithelial barrier defects and metabolic-induced immune dysregulation, while direct cellular and inflammatory responses to chemicals are covered in type VII. It is notable that several combinations of mixed types may appear in the clinical setting. The clinical relevance of the current approach for allergy practice will be conferred in another article that will follow this year, aiming at showing the relevance in clinical practice where various endotypes can overlap and evolve over the lifetime.

#### KEYWORDS

allergic diseases, EAACI position paper, hypersensitivity, nomenclature, pathophysiology and mechanism

## 1 | INTRODUCTION

The rapid growth of technology, including molecular diagnostics, omics technologies, genetic and epigenetic editing, nano-technologies, imaging and many more, generated in-depth knowledge of disease pathogenetic pathways (endotypes), allowing more detailed disease descriptions. The disease nomenclature usage of this new information set allowed a shift from a mere pathomechanistic approach, with symptoms and organ dysfunction as the primary descriptors, to recognition of a more established network of immunological and metabolic pathways described by various biomarkers, ideally validated. New disease endotypes, defined by distinct pathophysiological mechanisms, are now described for asthma,<sup>1-4</sup> allergic rhinitis (AR),<sup>5-7</sup> chronic rhinosinusitis (CRS),<sup>8,9</sup> atopic dermatitis (AD),<sup>10,11</sup> and food,<sup>12</sup> venom<sup>13</sup> and drug allergy.<sup>14</sup> However, the current disease taxonomy does not cover this information, so categorising diseases becomes more complicated, thus necessitating revision. A new nomenclature was needed describing diseases based on a combination of their intrinsic biology and traditional 'signs & symptoms', leading to a better understanding of aetiology, mechanisms, prevention, diagnosis and treatment. It should also be flexible, allowing easy incorporation of subsequently developing evidence into existing knowledge, and ideally, its implementation into daily practice should be effortless.

During the last two decades, high throughput technologies and analysis of multi-omics datasets have significantly contributed to increasing the resolution in identifying the triggers and pathomechanisms of diseases.<sup>15</sup> In addition, the introduction of precision medicine, the concepts of disease endotypes, genotypes, theratypes and regiotypes have helped to stratify patients based on disease mechanisms to optimize the management of allergic diseases.<sup>16-18</sup>

Artificial intelligence (AI) and machine learning have emerged as powerful tools for analysing complex datasets delivered by the new precision diagnostic tools, identifying patterns that may not

be readily apparent to human researchers. In the context of allergic diseases, AI can potentially support unbiased patient characterization based on their endotypes and more accurately predict responders and non-responders to targeted interventions or immune-modulating therapies. By uncovering novel biomarkers and identifying subgroups of patients with distinct immunological profiles, AI can facilitate the development of personalized treatment strategies (biologicals and small molecules/allergen immunotherapy and other immune-modulatory interventions), ultimately improving patient outcomes and achieving disease modification and targeted prevention.<sup>19</sup>

### 1.1 | Historical perspective

The term 'allergy' was first coined by Clemens von Pirquet in 1906. He derived the word from the Greek words 'allos' (meaning 'other' or 'different') and 'ergon' (meaning 'work' or 'reaction'). An allergy is an unexpected abnormal or exaggerated reaction to an exogenous stimulus involving the immune system.<sup>20</sup>

The term 'atopy' and the concept of atopic diseases were first proposed by the physicians Arthur F. Coca and Robert A. Cooke in the early 20th century. Atopy (or atopic) is a term derived from the Greek word 'topos', which means 'place', 'atopos' meaning 'out of place' or 'strange'. In 2001, Johansson et al. defined atopy as a personal or familial tendency to develop asthma, rhinoconjunctivitis or dermatitis due to sensitization to allergens. Individuals with atopy tend to have higher levels of immunoglobulin E (IgE) antibodies.<sup>21</sup>

The designation 'hypersensitivity' was introduced first in 1951 by Phillip Gell and Isabel Hinde referring to tuberculin reaction.<sup>22-24</sup> In 1963 Gell together with Robin Coombs defined hypersensitivity as an undesirable, uncomfortable or damaging response that arises from an overreaction of the adaptive immune response. It encompasses both allergies, which are triggered by external stimuli, and

autoimmunity, which arises from intrinsic stimuli. Typically, hypersensitivity reactions are a secondary immune response occurring in a host with a pre-sensitized (immune) state. According to Gell and Coombs, hypersensitivity reactions were classified into four types: type I: immediate (IgE-mediated), type II: cytotoxic (antibody and Fc receptor-mediated, cellular), type III: immune complex-mediated and type IV: delayed-type (T-cell-mediated).<sup>25</sup>

In 2001, the European Academy of Allergy and Clinical Immunology (EAACI) nomenclature task force led by Johansson et al. published the nomenclature for allergy. This document divides hypersensitivities into the following categories: IgE-mediated reactions that include atopic, and non-atopic conditions (insect sting, food allergy, drugs, helminths); non-IgE-mediated disorders, which are cell-mediated reactions that involve T-lymphocytes (contact dermatitis), IgG-mediated (allergic alveolitis) and other immune cells, for example, eosinophils (gastroenteropathy); non-allergic reactions that do not have the immune mechanisms involved.<sup>21,26</sup>

In the 1990s, several scientists translated the Mossman and Coffman murine Th1-Th2 model to human allergy and asthma.<sup>27-29</sup> In the late 1990s Werner Pichler proposed a further subdivision of type IV hypersensitivity reactions, based on the key cells, cytokines and chemokines. Type IVa (tuberculin reaction) are those reactions where monocytes (MOs) and macrophages (M $\phi$ ) are preferentially activated and recruited (with the subvariant of a granulomatous reaction); type IVb, where eosinophils and T helper lymphocytes type 2 (Th2) cells are preferentially activated and recruited; type IVc is mediated by cytotoxic functions of CD8<sup>+</sup> T-cells; and type IVd, where neutrophils (NEU) are preferentially activated and recruited.<sup>30-32</sup> Due to the current understanding that CD8<sup>+</sup> T-cells can be very diverse and analogous to CD4<sup>+</sup> T-cell subsets: CD8<sup>+</sup> subset 1 (Tc1), CD8<sup>+</sup> subset 2 (Tc2), CD8<sup>+</sup> subset 17 (Tc17), CD8<sup>+</sup> regulatory subset (-Treg), we have further modified the concept in this article – see below.

## 1.2 | Advances in immunology and allergy research

Given the advances in the understanding of immune mechanisms and novel therapeutic options, this EAACI Position Paper provides a necessary update of previous EAACI and World Allergy Organization nomenclature. The new nomenclature of allergic diseases is described in [Figure 1](#).

In brief, the novel concept described in this article is based on the understanding of distinct characteristics and functions of type 1, type 2 and type 3 complex immune responses and tissue-driven responses, which play crucial roles in immune-related disorders such as allergies or autoimmunity. *Type 1 immune responses (T1)* are directed towards intracellular pathogens such as *Mycobacterium tuberculosis* or viruses. CD4<sup>+</sup> T cells (Th1), type 1 innate lymphoid cells (ILC1), natural killer cells (NK), natural killer T cells (NK-T) and type 1 CD8<sup>+</sup> cytotoxic lymphocytes (Tc1) detect and kill infected cells and their contents. Interferon-gamma (IFN- $\gamma$ ) is the main effector cytokine, and IgG1, IgG2 and IgG3 are the main antibodies. These cells

interact among each other and tissue cells through the activation of M $\phi$  and NEU to eliminate intracellular pathogens. *Type 2 immune responses (T2)* protect against helminths (large protozoan infections), poisons and venoms. Key players are Th2, ILC2 and Tc2 cells, IgE and effector cells such as basophils, eosinophils and mast cells (MCs) with interleukin (IL)-4 and IL-5, IL-9, IL-13 as the main effector cytokines.<sup>33</sup> *Type 3 immune responses (T3)* are directed against extracellular bacteria and fungi characterized by Th17 cells, ILC3 and Tc17, with IL-17 as the main effector cytokine and NEU as the primary effector cells.<sup>34</sup> Mutations in involved genes, defects and deviation of these immune responses may lead to immune deficiencies, autoimmunity, cancer, abortions and allergies.

The modern definition of allergy should reflect the pathophysiological complexity of these conditions. They include conditions caused by hypersensitivity of the immune system elicited by otherwise harmless environmental substances. Although classically, the mechanisms of allergies are associated with T2 responses, recent discoveries showed endotypes of allergic diseases related to T1 or T3-driven activation pathways,<sup>34-36</sup> which were originally assumed to be involved in a variety of immune-mediated diseases including autoimmunity characterized by mechanisms distinct from allergies.<sup>37</sup>

Allergy is an abnormal or exaggerated reaction to exogenous stimuli which involves various types of hypersensitivity reactions engaging antibodies, immune cell-mediated, tissue-driven or metabolic mechanisms resulting in the development of respiratory, skin, eye, gastrointestinal and other symptoms, including anaphylaxis. Anaphylaxis is a serious systemic allergic reaction that is usually rapid in onset and is sometimes fatal.<sup>38,39</sup> The term atopy, although deeply rooted, has limited use today as it is based mainly on the symptomatic definition of diseases and does not represent the current understanding of the pathophysiology.

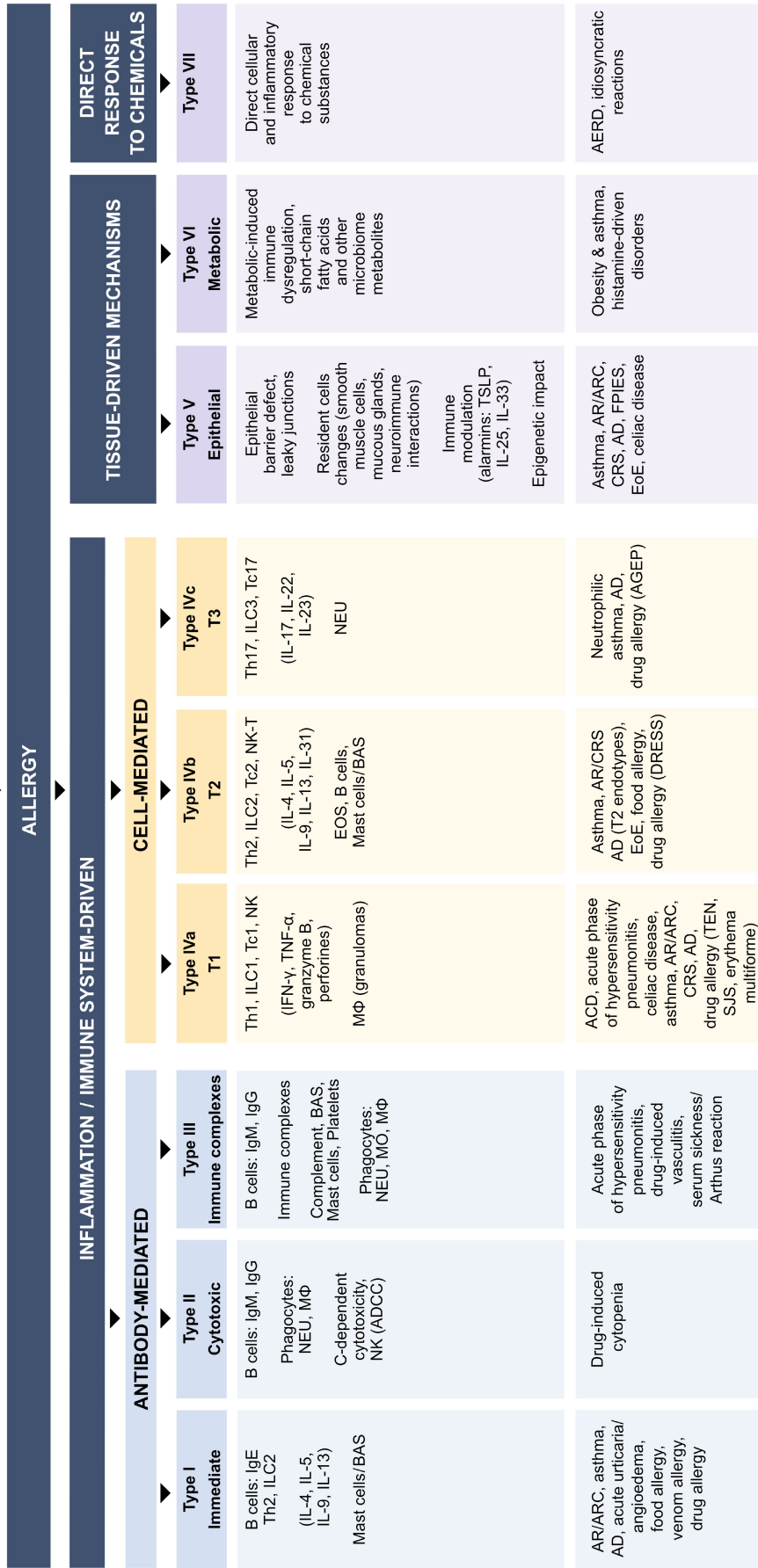
## 2 | MECHANISMS OF THE MOST IMPORTANT ALLERGIC DISEASES

### 2.1 | Type I or immediate response

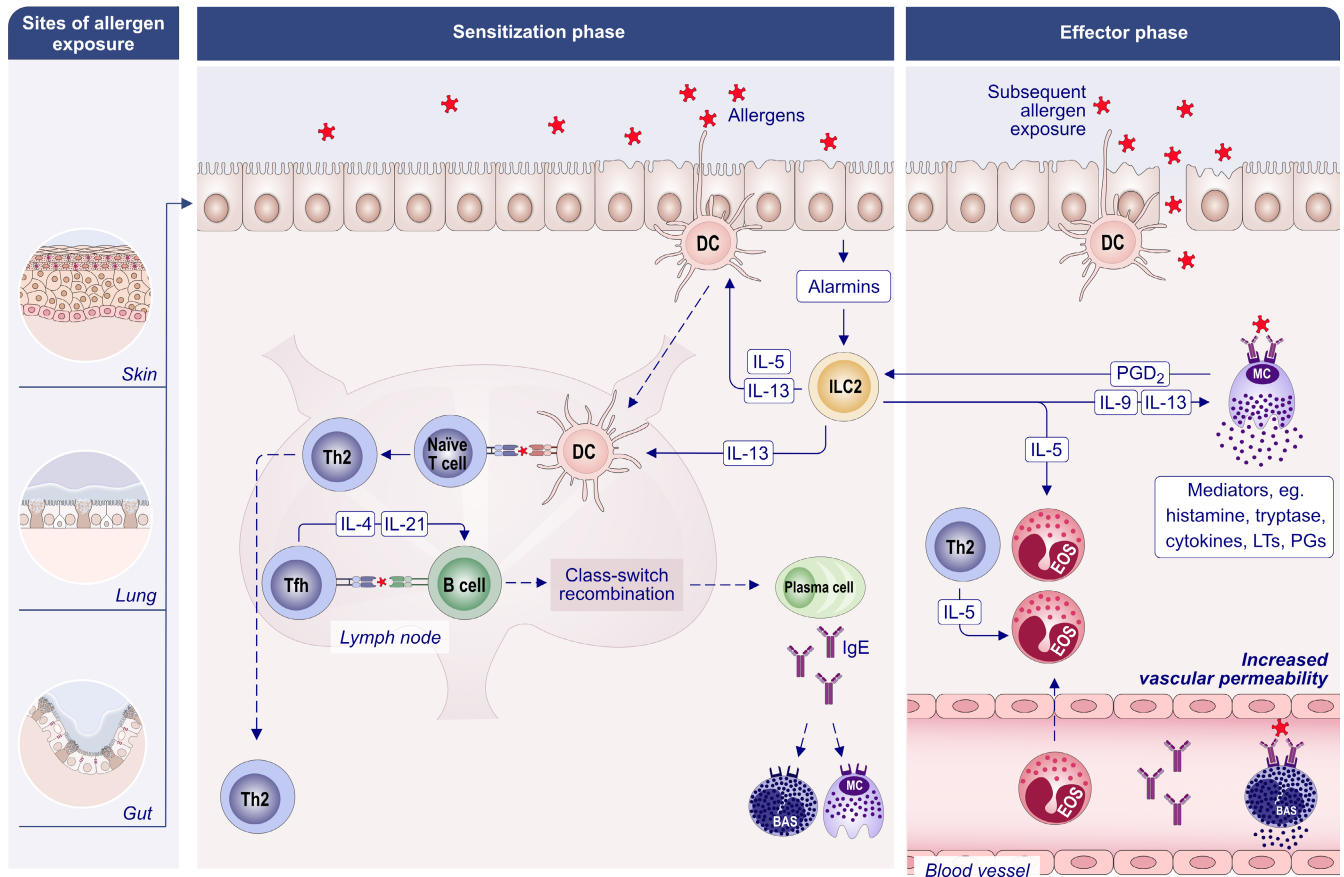
Type I, IgE-dependent reactions occur in patients with AR, allergic rhinoconjunctivitis (ARC), asthma, AD, acute urticaria/angioedema, food, venom and drug allergy ([Figure 2](#)).<sup>40</sup>

Classical allergens initiating type I hypersensitivity are pollens (trees, grasses and weeds), house dust mites (HDM), mould spores, cockroaches, animal dander, saliva and urine (e.g. cats, dogs, hamsters, guinea pigs), insect venoms (e.g. bees, wasps, ants), foods (e.g. peanuts, tree nuts, milk, eggs, fish, shellfish, soy, wheat, fruits, vegetables), latex (e.g. gloves, balloons and condoms) and drugs (e.g. penicillin and other beta-lactam antibiotics, serums and vaccines, insulin, monoclonal antibodies and other protein medications).

Type I response includes two phases. The *sensitization phase* depends on T2 cell signals (related to the hypersensitivity type IVb-described below) which regulate allergen-specific immunoglobulin E



**FIGURE 1** New nomenclature of allergic diseases. Hypersensitivity refers to an undesirable, uncomfortable or damaging response that arises from a tissue cell dysfunction or immune system overreaction. Allergy is an abnormal or exaggerated reaction to exogenous stimuli which involves various types of hypersensitivity reactions engaging antibodies, immune cell-mediated, tissue-driven or metabolic mechanisms resulting in the development of respiratory, skin, eye, gastrointestinal and other symptoms, including anaphylaxis. ACD, allergic contact dermatitis; AD, atopic dermatitis; ADCC, antibody-dependent cellular cytotoxicity; AERD, aspirin-exacerbated respiratory disease; AGEF, acute generalized exanthematous pustulosis; AR, allergic rhinitis; ARC, allergic rhinoconjunctivitis; B, B lymphocytes; BAS, basophil; CRS, chronic rhinosinusitis; DRESS, severe drug reaction with eosinophilia and systemic symptoms; EoE, eosinophilic oesophagitis; EOS, eosinophil; FPIES, food protein-induced enterocolitis syndrome; IFN-γ, interferon-gamma; Ig (E, G, M), immunoglobulin (type E, G, M); IL, interleukin; ILC1/2/3, innate lymphoid cells type 1/2/3; MO, monocyte; MΦ, macrophage; NEU, neutrophils; NK, natural killer cell; NK-T, natural killer T cell; SJS, Stevens-Johnson syndrome; T1/T2/T3, type 1/2/3 immune response; Tc1/2/17, T cytotoxic lymphocyte type 1/2/17; TEN, toxic epidermal necrolysis; Th, T helper lymphocytes; TILSP, thymic stromal lymphopoietin; TNF-α, tumour necrosis factor-alpha.



**FIGURE 2** Mechanisms of type I hypersensitivity in AR, ARC, asthma, AD, acute urticaria/angioedema, food, venom and drug allergy. The allergen is deposited on the epithelial cells, in the respiratory tract, gut or skin. The sensitization phase occurs after the first contact with the allergen, APCs, for example, DCs, present the antigen to the naïve Th. ILC2 are activated by cytokines released by epithelial cells (called alarmins), such as IL-25, IL-33 and TSLP. Upon activation, they produce large amounts of type 2 cytokines, including IL-5, IL-9 and IL-13, further supporting the T2-cell response. Tfh help B cells to mature and produce high-affinity sIgE. MC and BAS possess the high-affinity receptor for the Fc fragment of sIgE (FcεRI) and are coated with sIgE, thus concluding the sensitization phase. The effector phase occurs upon subsequent exposure to the same allergen. The allergen crosslinks sIgE bound to MC and BAS, triggering degranulation. MCs are located in various tissues throughout the body, while BAS circulate in the blood. Preformed mediators inside MC and BAS, like histamine, induce symptoms upon release into the microenvironment, like vasodilation, bronchial muscle contraction and increased mucus secretion. Eosinophils play a significant role in the delayed allergic response and the persistence of inflammation, engaging mechanisms related to type IVb hypersensitivity. Therefore, the mutual interaction between type I and IVb-related processes is vital to both the sensitization and the chronic phase. Asthma, AR, ARC and AD endotypes can show T2-type cytokine overexpression (IL-4, IL-5 and IL-13) and high serum sIgE levels. Food/venom/drug allergy can be induced directly by a trigger with a potentially life-threatening anaphylactic reaction. Acute urticaria/angioedema can be induced by allergens (e.g. foods, medications, insect bites or stings). B, B lymphocyte; BAS, basophil; DC, dendritic cell; EOS, eosinophil; IL, interleukin; ILC2, type 2 innate lymphoid cell; LT, leukotrienes; MC, mast cell; PG(D<sub>2</sub>), prostaglandin (D<sub>2</sub>); sIgE, allergen-specific immunoglobulin E; Tfh, T follicular helper cell; Th naïve/2, T helper lymphocyte naïve/type 2; TSLP, thymic stromal lymphopoietin.

(sIgE) production. The mechanism involves a complex interplay between the adaptive and innate immune systems. The process begins when an individual is exposed to an allergen for the first time. The allergen is internalized by antigen-presenting cells (APCs) such as dendritic cells (DC), B lymphocytes (B) and Mφ, which process and present the allergen peptides on their surface, linked to major histocompatibility complex class II (MHC class II) molecules to naïve T cells. DCs are shown to be the strongest activator of naïve T cells. However, B cells and Mφ can also contribute to naïve T-cell differentiation. Specific cytokines produced by APCs can vary depending on factors such as the nature of the antigen/allergen they encounter,

the local cytokine environment and the activation state of the APCs themselves.<sup>41</sup> DCs surface molecules, secreted metabolites and cytokines promote the activation and differentiation of naïve T-cells into various immune cell subsets, such as Th1, Th2, Th17, Tc1, Tc2, Tc17 and regulatory T cells. DCs do not secrete a typical type 2 immune response polarizing cytokines. Early IL-4 is produced by MCs and basophils, while ILC2s amplify and sustain the response. They are activated by cytokines released by epithelial cells (alarmins), such as IL-25, IL-33 and thymic stromal lymphopoietin (TSLP).<sup>42</sup> ILC2s can also be directly activated by environmental toxins.<sup>43</sup> Upon activation, ILC2 produce large amounts of type 2 cytokines, including IL-5,

IL-9 and IL-13, further supporting the T2 cell response, eosinophil recruitment and mucus production. There is limited data showing that ILC-2 produce IL-4, but this has not been fully elucidated.<sup>44,45</sup> This leads to the differentiation of naïve T cells into Th2 and Tc2 cells; IL-4 and IL-13 promote the immunoglobulin class-switch and thus the production of IgE by B cells and increase the tissue migration of Th2 cells. Additionally, ILC2-derived cytokines can promote local tissue repair and remodelling, contributing to chronic inflammation and tissue damage in cases of persistent allergen exposure.<sup>35,46</sup> T follicular helper cells (Tfh) are a subset of CD4<sup>+</sup> Th cells that play a critical role in the development and maturation of B cell responses, including the production of high-affinity antibodies. Tfh cells provide signals to B cells in the germinal centres, including cytokines (such as IL-4 and IL-21) and costimulatory molecules (such as CD40L). These signals help B cells undergo class-switch recombination, which results in the production of IgE.<sup>47</sup>

In many sensitized individuals, the clinical signature does not appear. The type I hypersensitivity response occurs due to deficient immune regulatory response, in ILC regulatory (ILCregs) cells, Tregs, Bregs and follicular regulatory T cells (Tfr).<sup>48-53</sup>

The *effector phase*: MCs and basophils express the high-affinity IgE receptor (FcεRI) for the Fc region of IgE. IgE, the least-abundant member of the antibodies, irreversibly binds to FcεRI on the surface of MCs and basophils, sensitizing these cells to the allergen. As a result, MCs, both mucosal and connective tissue subtypes, and basophils are coated with IgE. Upon subsequent exposure to the allergen, the allergen crosslinks two adjacent IgE on the cell surface, causing the cells to degranulate. The degranulation process releases pre-stored mediators such as histamine, heparin, proteases (e.g. tryptase) and some cytokines, as well as newly generated such as prostaglandins, leukotrienes and adenosine nucleotides. Upon activation, a MC can either slowly release (piecemeal degranulation) or rapidly release (anaphylactic degranulation) mediators or cytokines and chemokines that induce inflammation, from storage granules into the local tissue microenvironment.<sup>54</sup> These mediators cause the characteristic symptoms of an allergic reaction, including vasodilation, increased vascular permeability, smooth muscle contraction, stimulation of sensory nerves and mucus production.<sup>55</sup> IgE further upregulates the FcεRI on MCs allowing more IgE to bind to receptors and increase the mediator release upon allergen crosslinking. In addition, MCs can be activated by other non-IgE-related MC stimuli, described in section type VII.

Decreased levels of IgE and increased specific IgG4 can occur as part of the immune system's adaptive response to avoid allergic reactions after repetitive exposure to high concentrations of allergens, as in allergen immunotherapy (AIT). AIT can shift the immune response from a T2 cell-dominated response favouring IgE production to a T1 or Treg response, which supports the production of other immunoglobulins like IgA and IgG4.<sup>56</sup> Class-switching is associated with different cytokines, such as IFN-γ, IL-10 and tumour growth factor-beta (TGF-β). IgA and IgG4 can compete with IgE for allergen binding (blocking activity) without causing MCs or basophils activation. IgG4 can also suppress the production of IgE by B cells,

either directly or by inhibiting the activity of Th2 cells. IgA in mucosal secretions can bind to allergens, forming immune complexes that prevent the allergens from crossing the mucosal barrier and entering the circulation (immune exclusion). IgA can neutralize allergens' biological activity by binding to them and preventing them from inducing an immune response.<sup>57,58</sup>

Activation, migration and prolonged life span of eosinophils contribute to the late-phase allergic reaction and chronicity of inflammation, involving hypersensitivity type IVb mechanisms.<sup>59</sup> Thus reciprocal regulation of type I and type IVb-related mechanisms is crucial in allergy development during the sensitization and chronic phase of allergic disease.

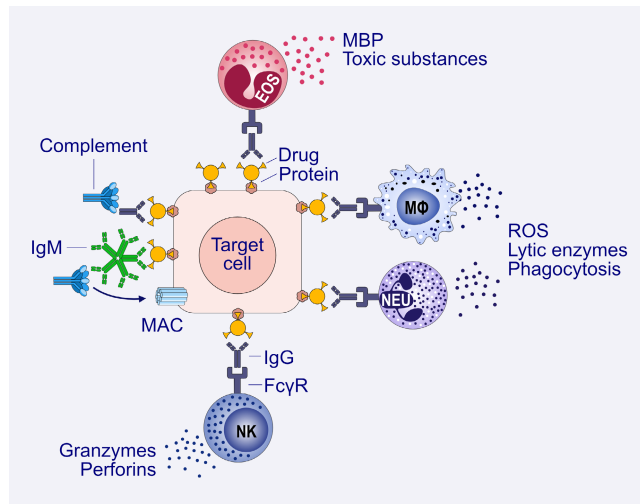
## 2.2 | Type II or antibody-mediated cellular cytotoxicity reaction

Type II reactions are typically drug-induced reactions which are considered a cause of allergic cytopenia. However, type II reactions are an essential pathogenetic event in several autoimmune diseases, such as immune thrombocytopenia, autoimmune haemolytic anaemia (AIHA), autoimmune neutropenia, Biermer's disease, Goodpasture syndrome, haemolytic disease of the foetus and the newborn (erythroblastosis fetalis), myasthenia gravis, pemphigus and transfusion reactions involving mismatched blood types.<sup>60-63</sup>

In a drug-dependent type II *allergic reactions*, a drug or its metabolite first binds to proteins on the cell membrane. Subsequently, anti-drug antibodies and drug-membrane protein complex bind and activate complement or are being bound by the Fc fragment gamma receptor (FcγR) on the effector cell such as an NK cell, eosinophil, Mφ or neutrophil, finally inducing cytolysis (Figure 3). The mechanisms of sensitization leading to the development of IgG are unclear and might potentially be due to molecular mimicry. Similarly, in type II *autoimmune reactions*, complexes of drug/anti-drug antibodies bind to the cell membrane on self-antigens and activate the complement or an effector cell via the Fc receptor, which results in cytolysis.

The primary antibodies involved in type II allergic reactions are IgG and IgM. The immunoglobulins damage cells by several mechanisms: (1) Activation of the classical way of the complement system generating the cytolytic membrane attack complex C5b-9. (2) Antibody-dependent cellular cytotoxicity (ADCC) mainly via NK cells and CD16 expressing CD8<sup>+</sup> T cells.<sup>64</sup> In ADCC, the IgG recognize antigens (drugs) bound to the target cell surface and then are bound by the FcγR on the effector cells. The effector cell releases cytotoxic substances, such as perforin and granzymes, that induce regulated cell death mechanisms, including apoptosis, necroptosis and pyroptosis.<sup>65,66</sup> (3) Opsonization of the drug-target cell by C3b and iC3b complement fragments or antibodies and phagocytosis by Mφ and NEU. (4) Activation of eosinophils through FcγR, and release, for example, major basic protein (MBP) or reactive oxygen species (ROS).<sup>59,67</sup>

The activation of the complement system and the recruitment of immune cells lead to the release of inflammatory preformed and



**FIGURE 3** Mechanisms of type II hypersensitivity, that include allergic cytopenia. The drug binds to the cell membrane proteins, and subsequently, an anti-drug antibody (IgM or IgG), bind to the complex drug-cell membrane. This leads to complement activation and cell membrane lysis. IgG can be bound by Fc $\gamma$ R on M $\phi$  and NEU, which activates phagocytosis, ROS and enzymes production. IgG can be also bound by Fc $\gamma$ R on EOS and cause the release of MBP or ROS. ADCC can be executed by NK or CD8<sup>+</sup> cells. The activation of complement and the recruitment of immune cells contribute to tissue damage. ADCC, Ab-dependent cellular cytotoxicity; EOS, eosinophil; Fc $\gamma$ R, Fc fragment gamma receptor; IgG/M, immunoglobulin class G/M; MAC, membrane-attack complex; M $\phi$ , macrophage; MBP, major basic protein; NEU, neutrophil; NK, natural killer cell; ROS, reactive oxygen species.

newly generated mediators, and proteolytic enzymes, contributing to further tissue damage.

### 2.3 | Type III or immune complex-mediated reactions

Type III *allergic reactions* include the acute phase of hypersensitivity pneumonitis (also referred to as extrinsic allergic alveolitis), drug-induced vasculitis, serum sickness and Arthus reaction. Type III reactions are associated with several *autoimmune diseases*, including systemic lupus erythematosus (SLE), rheumatoid arthritis (RA) and post-streptococcal glomerulonephritis.<sup>68</sup>

Type III allergic hypersensitivity reactions are mediated by IgM and IgG antibodies that bind soluble antigens, for example, drugs, venoms or other allergens, to form antigen-antibody complexes. As a result of the reduced clearance due to decreased function of the MOs activating system or increased production of antigen-antibody complexes (such as in chronic infections, autoimmune or neoplastic diseases), immune complexes deposit in various tissues throughout the body, such as small blood vessels, capillaries, joint synovium, kidney glomeruli and lung alveoli which are porous and permit the immune complexes to enter the tissues and cause inflammation. This leads to the extravascular activation of the complement system,

which releases chemotactic agents that attract NEU, causing inflammation and tissue damage (Figure 4).<sup>68</sup>

Complement activation stimulates a local inflammatory response, which causes the symptoms. In addition, complement-independent pathways involve the immune complexes with Fc $\gamma$ R on immune cells, such as NEU, undergoing pathologic frustrated phagocytosis. However, some research studies suggest that complement may play a minimal role in the actual process of allergic type III reactions.<sup>69</sup> Arthus reaction is only slightly reduced in mice with intact Fc signalling whose complement is decreased. In this case, MC degranulation appears to drive the entire reaction.<sup>70-72</sup> Complement, specifically the anaphylatoxin C5a, might indirectly drive the reaction by altering the ratio of activating to inhibitory Fc receptors on effector cells.<sup>73</sup> Subsequent aggregation of immune complex-related events may result in local fibrinoid necrosis with ischemia-aggravating thrombosis in tissue vessel walls.

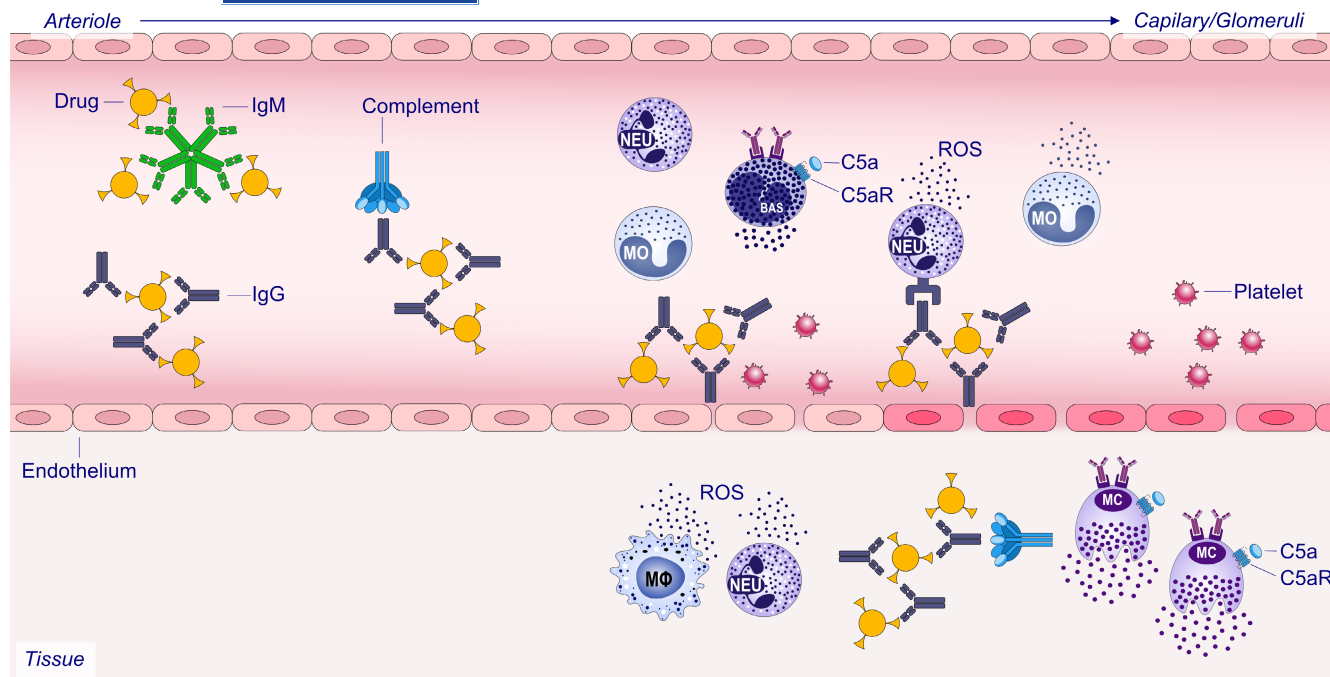
### 2.4 | Type IV or cell-mediated reactions

Memory T lymphocytes interacting with ILCs, NK-T cells, NK cells, NEU, eosinophils and M $\phi$  drive type IV reactions. Historically, these reactions were called delayed-type, due to the observation that symptoms develop several hours to days after exposure. Various T-cell subsets mediate Type IV responses through different specific pathways, displaying a high degree of heterogeneity reflecting the distinct phenotypic features of memory lymphocytes. Some disease mechanisms can be only explained by the cooperation of several subtypes of type IV hypersensitivity.

#### 2.4.1 | Type IVa – T1 immune response

Typical clinical manifestations of type IVa reaction are allergic contact dermatitis, the chronic phase of hypersensitivity pneumonitis (also referred to as extrinsic allergic alveolitis) and celiac disease. Type IVa reactions can also be essential for non-T2 endotypes of asthma, AR, CRS or AD. These mechanisms also explain non-immediate allergic reactions to drugs, which occur after the haptenization of the drug with a carrier protein.<sup>74</sup>

Type IVa reactions are considered to be T1 responses, mediated by memory Th1 and Tc1 cells, which acquire their phenotype upon exposure to IL-12, IL-23 and IFN- $\gamma$  provided by APC.<sup>75</sup> Th1 cells produce high amounts of IFN- $\gamma$ , lymphotoxin and tumour necrosis factor-alpha (TNF- $\alpha$ ), which contribute to many disease mechanisms, including granuloma formation, synthesis of IgG1 and IgG3 by B cells and the activation of T-cell cytotoxicity.<sup>76</sup> The memory immune response in type IVa reactions is amplified by innate immune cells, including ILC1 and classically-activated M $\phi$  (M1 M $\phi$ ), among others.<sup>77</sup> Activated M $\phi$  release various inflammatory mediators, such as ROS, proteases and pro-inflammatory cytokines, contributing to tissue damage at the site of antigen exposure. The tissue damage leads to the clinical manifestations of type IVa hypersensitivity, which can vary depending on the specific antigen and the affected tissue (Figure 5A).



**FIGURE 4** Mechanisms of type III hypersensitivity, including the acute phase of hypersensitivity pneumonitis, drug-induced vasculitis, serum sickness and the Arthus reaction. These reactions involve the formation of antigen–antibody complexes through the interaction of IgM and IgG antibodies with soluble antigens (e.g., drugs or venoms). These immune complexes are deposited in various body tissues, including blood vessels, joint synovium, kidney glomeruli and lung alveoli, due to impaired clearance by the M $\phi$  activating system. This triggers the complement system and increases vascular permeability, leading to cell chemotaxis. Recruiting NEU and MO cause inflammation through the release of enzymes and RO. BAS and MC release inflammatory mediators through binding the complement C5a fragment by C5aR. A cascade of events may ultimately lead to local fibrinoid necrosis, thrombosis and vasculitis. BAS, basophil; C5a(R), complement component 5a (receptor); MO, monocyte; MC, mast cell; M $\phi$ , macrophage; NEU, neutrophil; IgG/M, immunoglobulin class G/M; ROS, reactive oxygen species.

In addition, in T2-dependent asthma or AD, (hypersensitivity type VIb), after migrating to the bronchi or skin, Th2 cells can change their phenotype to produce T1 effector cytokines: IFN- $\gamma$ , TNF- $\alpha$  and express Fas-ligand and other apoptotic death signals that can induce bronchial epithelial or keratinocyte apoptosis followed by remodelling (Figure 5B).<sup>78,79</sup>

CD8<sup>+</sup> cytotoxic memory Tc1 cells also engage in type IVa reactions, especially in non-immediate allergic reactions to drugs.<sup>74</sup> Memory cytotoxic T cells usually differentiate upon exposure to IFN- $\gamma$  released by APC and Th1 cells.<sup>80</sup> Tc cells produce high amounts of IFN- $\gamma$ , mediating many inflammatory mechanisms.<sup>81</sup> The activation of memory Tc cells differs from that of memory Th cells.

**FIGURE 5** Mechanisms of type IVa hypersensitivity, including (A) ACD, chronic phase of hypersensitivity pneumonitis and celiac disease, (B) but also including non-T2 endotypes of asthma or AD. APC presents antigen/hapten to Th1 memory cells, which acquire their phenotype upon cytokine exposure, leading to the activation, proliferation and production of IFN- $\gamma$  and TNF- $\alpha$ . These cytokines recruit and activate immune cells, leading to inflammation and tissue damage. M $\phi$  are producing ROS, Tc and NK cells release granzymes and perforins. Innate immune cells, especially ILC1s, amplify the response by producing a large amount of IFN- $\gamma$ . Clinical manifestation of type IVa reaction is typical in ACD, where the triggering hapten is a small molecule which needs to associate with a host protein (e.g., an epidermal protein) to become immunogenic in a process called harmonization. In the intestine, celiac disease is mediated by gliadin-specific Th1 cells inducing intestinal inflammation upon the intake of wheat and other cereals. Chronic intestinal inflammation with damage of villi is associated with the generation of IgA and IgG antibodies against tissue proteins: anti-tissue transglutaminase Ab (tTG-IgA), anti-endomysial Ab (EMA-IgA), anti-deamidated gliadin peptides Ab (DGP-IgA and DGP-IgG), which turns the disease into a mixed allergic-autoimmune condition. The chronic phase of hypersensitivity pneumonitis is directed against airborne allergens that mediate inflammation in the lung parenchyma, ultimately leading to lung granuloma formation and scarring (fibrosis) of the lung tissue. (B) Th2 cells that migrate to the asthmatic bronchi, nasal mucosa or atopic skin often skew to additionally producing T1 effector cytokines: IFN- $\gamma$ , TNF- $\alpha$  and Fas-ligand (death signals) that can induce bronchial epithelial or keratinocyte apoptosis followed by remodelling. The CD8<sup>+</sup> T cells (Tc1), which respond to viral infections, can contribute to tissue inflammation and remodelling. Viruses activate Tc1 cells, which produce IFN- $\gamma$ , granzyme, etc. and induce airway hyperreactivity. ACD, allergic contact dermatitis; AD, atopic dermatitis; DC, dendritic cell; IFN- $\gamma$ , interferon-gamma; IgA/G, immunoglobulin A/G; IL, interleukin; ILC1, type 1 innate lymphoid cell; M $\phi$ , macrophage; ROS, reactive oxygen species; Th naïve/1/2, T helper lymphocyte naïve/1/2 type; Tc, cytotoxic lymphocyte; TNF- $\alpha$ , tumour necrosis factor-alpha.



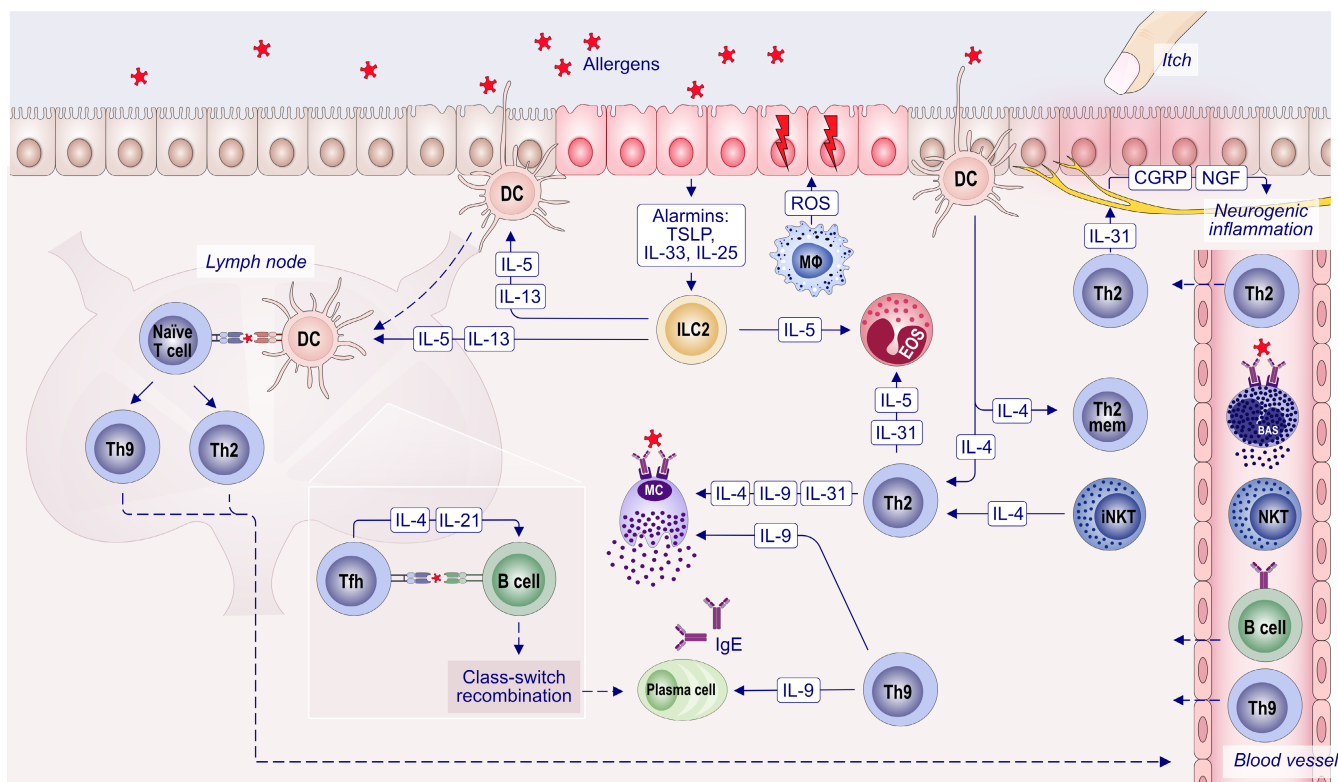


the lysis of the cell expressing the antigen in the context of MHC-I molecules.<sup>80</sup> In addition, TNF- $\alpha$ , Fas-ligand, TNF-like weak inducer of apoptosis (TWEAK) and TNF-related apoptosis-inducing ligand (TRAIL) signalling play a role in tissue injury, particularly epithelial cell apoptosis.<sup>83,84</sup> It has been shown that CD8<sup>+</sup> T cells, which play a crucial role in the antiviral immune defence can also trigger chronic allergic inflammation and remodelling. Rhinovirus, respiratory syncytial virus (RSV), influenza virus, parainfluenza virus, human metapneumovirus or coronaviruses (Sars-Cov-2) activate Tc1 cells, which

produce IFN- $\gamma$ , granzyme etc., leading to tissue damage and can also induce airway hyperreactivity.

## 2.4.2 | Type IVb – T2 immune response

The most characteristic expression of a type IVb hypersensitivity reaction can be observed in the classical allergic reaction with chronic airway inflammation in AR, CRS, asthma and AD (T2 endotype), food



**FIGURE 6** Mechanisms of type IVb hypersensitivity, including AR, AD, chronic rhinosinusitis with nasal polyposis and asthma (T2 endotype), but also EoE and food allergy. In Type IVb hypersensitivity reactions, Th2 cells play a central role, driven by cytokines such as IL-4, IL-13, IL-5, IL-9 and IL-31. These cytokines stimulate B cells to class switch to IgE (IL-4 and IL-13) and mediate eosinophilia (IL-5), causing inflammation and tissue damage. IL-31 mainly produced by Th2 cells, activates IL-31 receptors on sensory neurons, which release CGRP and NGF causing neurogenic inflammation and itch. Th9 cells, which differentiate with IL-4 and TGF- $\beta$ , contribute significantly to this response, enhancing sIgE synthesis and promoting MC growth. The response is further complicated by the ILC2 cells, MC and alternatively activated M $\phi$ . ILC2, DC and Th2 cells, activated by IL-25, IL-33 or TSLP, cooperate, producing cytokines and affecting epithelial barriers. They facilitate eosinophil and basophil recruitment and modulate APC function, contributing to the chronicity of Type IVb reactions. iNKT cells contribute to this response by producing IL-4 and IL-13, which induce alternative activation in M $\phi$  and promote inflammation. Eosinophils migrate to inflammatory sites, activate various cytokines and chemokines and release cytotoxic granules contributing to tissue damage, cell death and chronic inflammation. At the final stage when IgE synthesis is triggered, type IVb and I overlap. T2-high asthma is characterized by eosinophilic airway infiltrates and Th2-dependent cytokine overexpression (IL-4, IL-5 and IL-13). In AD, the most common endotype is characterized by high serum IgE levels and a strong association with other allergic diseases such as asthma and AR. Both IgE-dependent and IgE-independent pathways characterize mixed food allergy. Atopic manifestations arising from IgE-independent factors include delayed food-allergy-associated atopic dermatitis (6–48 h post-exposure) caused by the T2 cells (hypersensitivity type IVb), and eosinophilic gastrointestinal disorders, such as EoE. The oesophageal epithelium is the source of the IL-1 cytokine family members (IL-33, IL-36) and TSLP, involved in the balancing of pro- and anti-inflammatory responses. AD, atopic dermatitis; AR, allergic rhinitis; APC, antigen-presenting cell; B, B lymphocyte; BAS, basophil; CGRP, calcitonin gene-related peptide; DC, dendritic cell; EOS, eosinophil; EoE, eosinophilic oesophagitis; (s) IgE, (allergen-specific) immunoglobulins class E; ILC2, type 2 innate lymphoid cell; IL, interleukin; (i)NKT, (invariant) natural killer T cells; M $\phi$ , macrophage; MC, mast cell; Mf, macrophage; NGF, nerve growth factor; ROS, reactive oxygen species; Th0/2/9, T helper lymphocyte naïve/ type 2 or 9; T2, type 2 immune response; Tfh, T follicular helper cell; TGF- $\beta$ , tumour necrosis factor beta; Th mem, Th memory cells; TSLP, thymic stromal lymphopoietin.

allergy, eosinophilic oesophagitis (EoE) or protein-contact dermatitis. Th2 cells, ILC2, NK-T cells, eosinophils and a subset of M $\phi$  are the main players in the Type IVb – T2 immune response.

Type IVb reactions are mediated by Th2 cells, which acquire their phenotype upon exposure to IL-4, basophils or NK-T cells.<sup>85</sup> Th2 cells produce high amounts of IL-4, IL-5, IL-9, IL-13, IL-31 and eotaxins I-III. IL-4 and IL-13 are the key cytokines of type IVb hypersensitivity and induce a class switch to IgE in B cells by direct (from IgM) and indirect (from IgG1) mechanisms.<sup>86</sup> IL-13 is responsible for the tissue remodelling accompanying chronicity in type IVb hypersensitivity, whereas IL-5 mediates the bone marrow expansion of eosinophils, circulating eosinophilia and recruitment of eosinophils to the sites of inflammation and their survival in the tissues; eosinophils degranulate releasing their endogenous proteases into the microenvironment causing further tissue injury, chronic tissue damage and barrier disruption.<sup>87</sup> IL-31 is the main cytokine playing a role in itch. It is mainly produced by Th2 cells but also by M $\phi$  and DC. Its receptor is expressed on sensory neurons, epithelial cells or keratinocytes.<sup>88,89</sup> Th2 immune responses are often accompanied by allergen-specific Th9 cells, which differentiate upon exposure to IL-4 and TGF- $\beta$ .<sup>90</sup> Th9 cells can be considered substantial players in type IVb hypersensitivity. They produce IL-9, which enhances IL-4-mediated synthesis of IgE by B cells and is an important growth factor for MC precursors in the bone marrow, eosinophils and basophils.<sup>91</sup> The memory immune response in type IVb reactions is amplified by the activation of innate immune cells such as MCs, basophils, ILC2, eosinophils or alternatively-activated M $\phi$ , among others (Figure 6).<sup>92</sup>

ILC2 cells can produce type 2 cytokines, particularly IL-5, IL-13, IL-9 and amphiregulin to mediate T2 immune protection against helminths, causing tissue inflammation and tissue homeostasis. ILC2 are activated in response to IL-33 and/or IL-25 from epithelial cells. They crosstalk with T2 pathways, play a role in the recruitment of eosinophils and basophils and activate APCs that support a T2 response.<sup>93</sup> ILC2s, together with Th2 cells, open epithelial barriers via IL-13 and play a role in the resolving of tissue inflammation.<sup>94,95</sup>

MCs and basophils, and possibly ILC-2, provide the early source of IL-4 involved in Th2 cell differentiation.<sup>96,97</sup> In addition, IL-4 is produced by a unique subset of invariant natural killer T (iNK-T) cells (NK-T2), contributing to the activation of CD4<sup>+</sup> and CD8<sup>+</sup> T2 cells and the initiation and ongoing T2 inflammation via IL-4. In addition, a small fraction of IL-13-producing T2 NK and NK-T cells have been shown in the non-IFN- $\gamma$  secreting group.<sup>98</sup> IL-4 and IL-13 induce an alternative activation program in M $\phi$  that become suppressors of T1-linked cellular activities. A subset of M $\phi$  governs T2 functions at the interface of immunity, and tissue homeostasis and can produce IL-13.<sup>99</sup>

Eosinophils are the main players in all type IVb-T2 immune responses and contribute to chronic allergic inflammation. Mature eosinophils circulate in the blood and migrate to tissue sites of type IVb inflammation and helminth infection. Eosinophils are activated by various cytokines (such as IL-5) released by immune cells like Th2 and MC.<sup>100</sup> Eosinophils are also activated in response to chemokines like eotaxin-1 (CCL11), eotaxin-2 (CCL24), C-C motif

chemokine ligand 5 (CCL5 or RANTES), 5-hydroxy eicosatetraenoic acid and 5-oxo-eicosatetraenoic acid and certain leukotrienes like leukotriene B4 and MO chemoattractant proteins. IL-13 stimulates eosinophilic exit from the bone marrow. Activated eosinophils release cytotoxic granules containing proteins like major basic protein (MBP), eosinophil cationic protein, eosinophil-derived neurotoxin and eosinophil peroxidase.<sup>59</sup> Eosinophils can undergo cytolysis, where granules are directly exposed to the extracellular environment, which allows them to exert pronounced direct effects on surrounding tissues.<sup>101</sup> These proteins can cause tissue damage and contribute to the inflammation and symptoms associated with allergic reactions. Activated eosinophils produce extracellular traps (EET) that cause cellular damage due to their eosinophilic toxin content.<sup>102</sup>

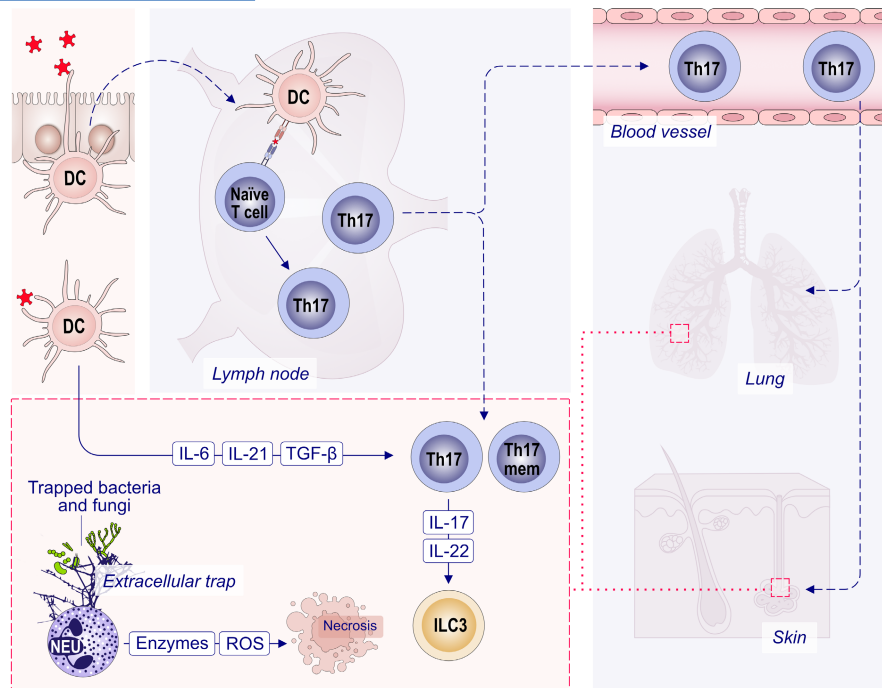
Type IVb and type I hypersensitivity overlap at the very final stage when IgE synthesis is triggered. T2 cells, through the released cytokines (IL-4, IL-13) induce IgE synthesis. However, the major type IVb-related effector mechanism involves eosinophil activation through IL-5. In addition, type IVb and type V hypersensitivity overlap in epithelial cell activation and opening of the epithelial barriers and drainage of the inflammation towards the bronchial lumen.<sup>95</sup>

### 2.4.3 | Type IVc – T3 immune response

Th17, Tc17, ILC3 and other IL-17A- and IL-17F-producing cells have been implicated in neutrophilic inflammation and the pathogenesis of AD and neutrophilic asthma.

In type IVc responses, Th17 cells, which belong to the helper T-cell lineage, produce IL-17 family cytokines that regulate innate effectors and orchestrate local inflammation by inducing the release of proinflammatory cytokines and chemokines capable of recruiting NEU and enhancing Th2 cytokine production. Memory Th17 cells acquire their phenotype upon exposure to IL-6, IL-21, IL-23 and TGF- $\beta$  provided by APCs. The main effector cytokines produced by Th17 cells are IL-17A, IL-17F, IL-21, IL-22 and granulocyte-M $\phi$  colony-stimulating factor.<sup>103</sup> IL-17A and IL-17F are produced by CD4<sup>+</sup> and CD8<sup>+</sup> T cells, gamma delta T cells and NK cells in response to IL-1 $\beta$  and IL-23. Their default role is protective immunity against fungi and bacteria by promoting antimicrobial peptide production, neutrophil recruitment and enhanced epithelial barrier function.<sup>104</sup> IL-17A and IL-17F activate ILC3 and stromal cells to produce IL-8, which recruits NEU to the sites of inflammation.<sup>105</sup> Thus, tissue infiltration by NEU is the hallmark of type IVc hypersensitivity. In addition to the 'respiratory burst' and enzyme release, which cause necrosis, neutrophil extracellular traps (NETs) can be associated with host damage. NETs are networks of extracellular fibres, primarily composed of DNA (Figure 7).<sup>106</sup> Similar mechanism, extracellular traps – EET, have eosinophils (see above – type IVb).

The T3 response can be amplified by innate immune cells, especially ILC3. Type IVc inflammation often accompanies type IVa



**FIGURE 7** Mechanisms of type IVc hypersensitivity, in the pathogenesis of AD, chronic rhinosinusitis with nasal polyposis and neutrophilic asthma. In type IVc hypersensitivity, Th17 cells and ILC3 play a key role by producing IL-17 family cytokines, which induce neutrophil recruitment and enhance Th2 cytokine production, leading to inflammation. Certain bacterial and fungal components can promote Th17 responses. These responses can result in tissue damage via 'respiratory burst', enzyme release or NETs. NETosis is an important part of anti-bacterial and -fungal response, by trapping and killing various pathogens. This mechanism is particularly relevant to conditions like AD and neutrophilic asthma. Normal serum IgE levels might characterize the AD endotype with a more diverse immune profile involving Th1, Th17 and Th22 cells. The skin barrier dysfunction in AD is more severe, and these patients are more susceptible to irritant contact dermatitis. AD, atopic dermatitis; DC, dendritic cell; IgE, immunoglobulin E; IL, interleukin; ILC3, type 3 innate lymphoid cell; NET, neutrophil extracellular trap; NET, neutrophil extracellular trap; NK, natural killer cell; ROS, reactive oxygen species; TGF- $\beta$ , tumour necrosis factor-beta; Th naïve/2/17/22, T helper lymphocyte naïve/2/17/22 type; Th17 mem, memory Th17.

reactions. However, in some pathologies, the activation of memory Th17 cells prevails.<sup>107</sup>

#### 2.4.4 | Other possible types of type IV hypersensitivity

The p-i concept (=pharmacological interaction with immune receptors) postulates that some drugs can bind directly and reversibly (non-covalently) to immune receptors and thereby stimulate the cells. A certain drug may bind to a particular T-cell receptor (TCR) or bind directly to a certain HLA-molecule, which would explain the striking HLA associations of some drug hypersensitivity reactions. This drug binding suffices - together with TCR interactions with the HLA - to stimulate the T-cell to secrete cytokines, increase, and exert cytotoxicity.<sup>108</sup>

[Correction added on 3 November 2023, after first online publication: In section 2.4.4 Other possible types of type IV hypersensitivity, the preceding paragraph and reference no. 108 have been added to this version.]

More subtypes of IV reactions could be described based on the driver effector T cells. These can include, for example, Th9 or Th22 cells.<sup>109,110</sup> IL-9 is the prototypic cytokine that influences various target cells such as T cells, B cells, MCs and airway epithelial

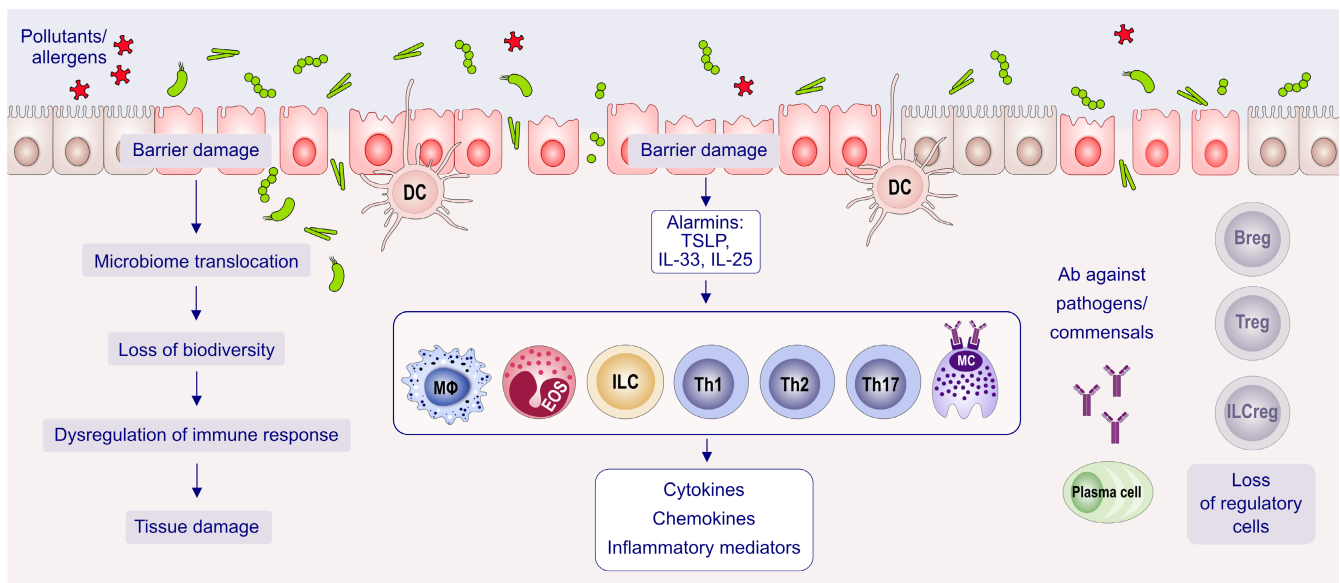
cells by activating members of the signal transducer and activator of transcription (STAT) proteins 1, STAT3 and STAT5. Th9 cells may promote immune tolerance in some models<sup>111</sup> and protect against parasitic infections,<sup>112</sup> they also trigger allergic inflammation and asthma,<sup>113</sup> which highlight their pleiotropic role in the immune system. CD4<sup>+</sup> T-cell subsets (Th17, Th9), MCs and ILC2s, can produce IL-9. Among other effects, IL-9 is a key cytokine for Th17 and Treg differentiation,<sup>114</sup> increases IL-4-mediated production of IgE and IgG by B cells,<sup>115</sup> and enhances the growth of bone marrow MCs and MC progenitors together with stem cell factor.<sup>116</sup> Th22 cells play a tissue-protective role at the early stages of asthma and AD, and are involved in tissue remodelling at the chronic phase.<sup>117-119</sup> The prototypic cytokine is IL-22. IL-22 primarily targets nonhematopoietic epithelial and stromal cells promoting proliferation and playing a role in tissue regeneration. In addition, IL-22 regulates host defence at barrier surfaces. In contrast, a proinflammatory role of IL-22 in the skin has also been proposed, as the severity of AD is associated with increased levels of CD8<sup>+</sup> IL-22-secreting cells.<sup>120</sup>

#### 2.5 | Type V - epithelial barrier defect

In recent years, significant progress has been made in understanding the different phenotypes and endotypes of mucosal/cutaneous

inflammatory diseases such as chronic AR/ARC, CRS, AD, asthma or food protein-induced enterocolitis syndrome, EoE and celiac disease. This revealed that these conditions are not homogeneous diseases but are instead defined by a constellation of symptoms which may result from different pathological mechanisms.<sup>121</sup> In some cases, the inflammatory process appears to reflect altered barrier function of the skin or mucosa, rather than from a primary immune dysregulation.<sup>122</sup> The impairment of the epithelial barrier function facilitates the activation of the underlying immune system and subsequently leads to chronic inflammation. Barrier loss can result from defects in several essential components, including stratum corneum structural elements in skin, tight junction proteins in skin and mucosa, protective antiproteases, expression of antimicrobial products, transport of ions, protons, water or antimicrobial materials and other mechanisms. The activation of sensory nerves, which contributes to the development of allergic symptoms, is also associated with the loss of barrier (Figure 8).<sup>123</sup> Intestinal barrier dysfunction may also occur via mucus erosion through low fibre-containing nutrition/diet.<sup>124,125</sup> This accounts for the rationale to introduce type V hypersensitivity to point out the peculiarities of the pathological processes and due to their importance in the view of personalized and precision approaches to endotype and biomarkers characterization and rapidly developing biological treatment, especially with anti-alarmins.

Mutations in filaggrin, a keratin-binding protein essential for epidermal homeostasis, significantly predispose to AD, in individuals with and without IgE-sensitization to allergens.<sup>126</sup> It has been suggested that filaggrin mutations could exert a similar effect on mucosa and predispose to diseases like asthma.<sup>127,128</sup> The impairment of the epithelial barrier can also arise from inflammatory phenomena.<sup>95</sup> It has been shown that IL-13 derived from Th2 cells and ILC2s markedly disrupts epithelial tight junctions.<sup>94,129</sup> The activation and tissue migration of ILC2 and Th2 cells strongly depend on epithelial-derived alarmins, especially IL-33, IL-25 and TLPS.<sup>130</sup> Interestingly, the protease activity of some aeroallergens, like HDM, might account for the activation of airway epithelial cells, the release of IL-33, the stimulation of ILC2, the production of IL-13 and the ultimate disruption of epithelial tight junctions.<sup>131</sup> These linked phenomena can occur without lymphocytes or activation of adaptive immunity.<sup>95</sup> It has been shown that the tight junctions in the airway mucosa of patients with allergic rhinitis and asthma are also disrupted. This illustrates how the barrier defect can also occur in classical type IVb hypersensitivity.<sup>132</sup> These data demonstrate the intricate relationship between different hypersensitivity mechanisms in the skin and mucosa: epithelial integrity can be dampened by intrinsic defects, but inflammatory phenomena can also cause an impairment of the barrier function, which further activates the immune system.



**FIGURE 8** Mechanisms of type V hypersensitivity; include asthma, chronic AR/ARC, CRS, AD, FPIES, EoE and celiac disease. The epithelial barrier defect and microbial dysbiosis lead to dysregulation of the immune response, including extensive activation of T1, T2 and T17 responses combined with the loss of Treg, Bregs and ILCregs. Additionally, formation of sIgE to inhaled or ingested allergens, activation of Mφ, MC and BAS and release of proinflammatory cytokines, chemokines and inflammatory mediators (histamine, leukotrienes, ROS). The sequence of events eventually leads to tissue damage that can be seen in asthma, chronic AR/ARC, CRS, AD, FPIES, EoE and celiac disease. Immune response to opportunistic pathogens and commensals, for example, *Staphylococcus aureus* (microbiome translocation) leads to IgE antibody production against them. Ab, antibody; AD, atopic dermatitis; AR/ARC, allergic rhinitis/rhinoconjunctivitis; BAS, basophil; Breg, B regulatory cells; CRS, chronic rhinosinusitis; DC, dendritic cell; EOS, eosinophil; EoE, eosinophilic oesophagitis; IL, interleukin; ILC, innate lymphoid cell; ILCreg, ILC regulatory cells; MC, mast cell; Mφ, macrophage; FPIES, food protein-induced enterocolitis syndrome; ROS, reactive oxygen species; sIgE, allergen-specific immunoglobulins class E; Th1/2/17, T helper lymphocyte type 1/2/17; Treg, T regulatory cells; TSLP, thymic stromal lymphopoietin.

As an important factor in type V hypersensitivity, the direct involvement of environmental factors that directly disrupt epithelial barriers has been recently demonstrated in several models and human tissues.<sup>43</sup> Direct exposure to air pollutants, chemicals and other environmental factors in the exposome can disrupt the epithelial barriers and affect the microbiome and immune system. Many of the chemical agents found in common consumer products (including toothpaste, shampoo, detergents and processed foods), are known to damage these critical barriers, increasing permeability to bacteria, toxins, pollutants and allergens.<sup>133</sup> When epithelial barriers are disrupted (or 'leaky'), substances and microbes can pass into deeper tissues, where they normally do not belong and trigger an immune/inflammatory response that can initiate or aggravate many chronic inflammatory diseases via inflammasome pathways.<sup>134</sup> Epithelial barrier defects have been demonstrated not only in T2 responses but also in non-T2 responses in chronic rhinosinusitis with nasal polyposis (CRSwNP) and non-T2 asthma. Recent exposure studies in mouse models of eosinophilic lung inflammation in asthma and eosinophilic esophagitis development in response to sodium lauryl sulphate and detergents demonstrate that asthma and (EoE)-like inflammation start with only epithelial cell activation and barrier leakiness-induced with toxic substances.<sup>43,135</sup>

As a direct example of chemical cytotoxicity, individuals with leaky epithelial barriers exhibit local inflammation in their epithelial cells, referred to as 'epithelitis'. Epithelitis is the initial event that attracts proinflammatory cells to the damaged epithelial barrier area. It starts with environmental insults (pollutants and toxic substance exposure), viral infections and enzymes in allergens. Mainly the alarmins, IL-25, IL-33 and TSLP and numerous proinflammatory chemokines are released by the epithelial cells inviting the immune system, into the area, particularly the Type IVb and players of the T2 response.<sup>122</sup>

Microbial dysbiosis takes place in areas of leaky inflammation epithelial barrier. A healthy microbiota on the surface of the mucosal barrier regulates numerous aspects of the barrier homeostasis. However, reduced biodiversity and alterations in the composition and metabolism of gut and skin microbiota are associated with various inflammatory conditions, including asthma, allergic diseases, inflammatory bowel disease, type 1 diabetes and obesity.<sup>136</sup> Dysbiosis refers to an imbalance in the microorganisms residing in our tissues, with microbial dysbiosis and bacterial translocation linked to the development and exacerbation of allergic and autoimmune diseases.<sup>137</sup>

## 2.6 | Type VI – metabolic-induced immune dysregulation

Concurrent with a general increase in the rates of obesity, the number of obese patients with asthma has also risen dramatically during the last few years.<sup>138</sup> Obesity is a distinguishing variable for clustering and classifying asthma subtypes. The obese asthmatic, more likely to be female with adult-onset asthma, and more likely

to become corticosteroid resistant, has a higher risk of being hospitalized and more frequently presents with severe disease.<sup>139</sup> Obesity can influence asthma by altering chest wall dynamics. It can influence inflammatory responses directly (e.g. via the release of inflammatory mediators from adipose tissue) or indirectly (e.g. due to the typical dietary changes associated with obesity, such as high levels of processed fats and low levels of fibre).<sup>140</sup> Increasing body mass index (BMI) is associated with increased levels of circulating inflammatory mediators and increased blood neutrophil and eosinophil counts.<sup>141</sup> Obesity is associated with increased levels of circulating serum acute phase reactants, ROS, chemokines and innate proinflammatory cytokines as well as those directly derived from adipose tissue (e.g. leptin),<sup>142</sup> but usually with no increase in serum cytokines associated with T-cell polarization, such as T<sub>H</sub>2 cytokines. While stressed and hypoxic adipocytes contribute to the pool of inflammatory mediators observed in serum, activated tissue M $\phi$  are particularly important to inflammatory responses and metabolic dysfunction associated with obesity.<sup>143</sup>

In obese asthmatics, there is an additive effect of asthma and obesity on increased release of pro-inflammatory mediators and airway (allergic) inflammation, as well as modification of the gut, nasal, oral and lung microbiome, closely linked with inflammatory responses.<sup>144</sup> While IL-5 levels are elevated in the bronchoalveolar lavages (BALs) of obese and non-obese asthmatics, only obese asthmatic BALs and lung tissue biopsies display increased activation of innate inflammatory pathways and enhanced activation of pathways associated with airway remodelling, which were not observed in the non-obese asthmatic lung. The obese asthmatic displays immune and inflammatory features associated with both asthma and obesity. Metabolic dysregulation in diabetes and obesity is linked to epithelial barrier leakiness. Immune cells activated at leaky barrier sites, particularly in the gut, can migrate to distant organs, causing inflammation in those areas. Moreover, increased inflammatory mediators in the circulation, namely, 'circulating microinflammation', consisting of acute phase reactants, chemokines and cytokines, can be detected and cause a metabolic burden.<sup>145</sup> In parallel with metabolic changes, perturbation of the intestinal microbiota, together with a persistent low-grade inflammatory response in the gut and fat tissue, is observed in obesity.<sup>146</sup>

The role of the microbiome and bacterial-derived mediators such as histamine in metabolic-induced immune dysregulation has been postulated. The imbalance in the gut microbiome, known as dysbiosis, can lead to a deviated immune response and increase the risk of chronic disease, including allergies or autoimmunity. Histamine, a mediator of inflammation, has been shown to regulate the immune response and is synthesized by certain bacteria, such as *Lactobacillus* and *Escherichia*, in the gut microbiome.<sup>147,148</sup> Histamine type 2 receptor plays a crucial role in modulating Th2- and Treg-cell activity.<sup>149</sup> Multiple novel immune modifying metabolites (e.g. tryptophan metabolites, short-chain fatty acids) have been recently identified and dysregulated secretion of these immunomodulators may also contribute to the development of allergies.<sup>150</sup>

## 2.7 | Type VII – direct cellular and inflammatory response to chemical substances

Type VII reactions occur in patients with AR, ARC, asthma, AD, acute urticaria/angioedema and drug allergy.

Idiosyncratic reactions include cross-reactive hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs). These reactions include at least three different phenotypes depending on the presence or absence of underlying respiratory or cutaneous disease: NSAIDs-exacerbated respiratory disease, in patients with rhinitis and/or asthma with or without nasal polyposis; NSAIDs-exacerbated cutaneous disease, in patients with underlying chronic spontaneous urticaria; and NSAIDs-acute urticaria/angioedema, in otherwise healthy individuals.<sup>151</sup> Recently, other phenotypes consisting of the simultaneous presence of cutaneous and respiratory symptoms after intake of NSAIDs have been extensively described.<sup>152</sup> The underlying mechanism in these reactions was linked to the inhibition of cyclooxygenase (COX)-1 and the release of eicosanoid mediators in susceptible individuals and has also been recently proposed for NSAID-induced acute urticaria/angioedema.<sup>153</sup>

Aspirin-exacerbated respiratory disease (AERD), also called NSAID-exacerbated respiratory disease (N-ERD) and previously called Samter's disease, is a chronic inflammatory condition characterized by the triad of asthma, recurrent nasal polyps and hypersensitivity to aspirin and other NSAIDs.<sup>154</sup> Arachidonic acid is a fatty acid that serves as a precursor for the synthesis of various eicosanoids, including prostaglandins (PG) and leukotrienes.<sup>155</sup> In AERD, there is an imbalance in the metabolism of arachidonic acid, leading to an overproduction of cysteinyl leukotrienes and a decrease in anti-inflammatory PG.<sup>156</sup> Aspirin and other NSAIDs inhibit the COX-1 and COX-2 enzymes responsible for synthesizing PG and leukotrienes. This inhibition further exacerbates the imbalance, resulting in a more pronounced inflammatory response. Cysteinyl leukotrienes (LTC<sub>4</sub>, LTD<sub>4</sub> and LTE<sub>4</sub>) are potent inflammatory mediators crucial in AERD pathogenesis.<sup>157</sup> They cause bronchoconstriction, increased vascular permeability, mucus production and recruitment of inflammatory cells. Platelets are more easily activated in AERD patients, releasing various inflammatory mediators, such as thromboxane A<sub>2</sub> and platelet-activating factor. The imbalance between Th2 cells and regulatory Tregs causes Th2 cells to secrete cytokines that promote eosinophilic inflammation, while Tregs help suppress excessive immune responses.<sup>158</sup> Airway epithelial remodelling in AERD is more than goblet cell metaplasia, resulting in a network of dysregulated inflammatory pathways and epithelial MC changes that sustain T2 inflammation.<sup>159</sup>

Examples of pharmacological interactions are represented by reactions mediated by G protein-coupled receptors (GPCR) expressed in MCs. A novel GPCR known as Mas-related GPCR X2 (MRGPRX2) has been recently described, which is activated in an Ab-independent manner by a range of cationic ligands (secretagogues) and is thought to be responsible for anaphylactoid reactions. These ligands include inflammatory peptides and drugs associated with allergic-type reactions, such as non-depolarizing neuromuscular blockers (atracurium, mivacurium, tubocurarine and rocuronium)

and fluoroquinolones (ciprofloxacin, levofloxacin, moxifloxacin and ofloxacin), among others. Anti-fungal antibiotics, aminoglycosides and sulphonamides induce MC degranulation through this receptor.<sup>160</sup> In addition, MRGPRX2 expression is increased in the skin of patients with severe chronic urticaria.<sup>161</sup> Representative members of each evaluated drug group led to the release of histamine, TNF, PGD<sub>2</sub> and b-hexosaminidase from MCs in leukocyte adhesion deficiency type 2 subjects.<sup>162</sup>

In addition, MCs can be activated by various compounds through their associated G-protein coupled receptors (e.g. morphine, radio-contrast media) by ion channels or, in the case of mucosal MCs, by hyperosmolar stimuli (as in exercise-induced asthma or by physical injury through pattern recognition receptors (PRR) that are triggered by damage-associated molecular patterns, by microbial pathogens through PRRs for pathogen-associated molecular patterns. Furthermore, complement components can also activate membrane receptors on MCs to exert various functions as well.<sup>163,164</sup>

## 3 | CONCLUSIONS

The dissemination and acceptance of the new nomenclature of allergic diseases proposed herein is important for the progress in the entire field. This approach is based on disease mechanisms and endotypes rather than phenotypes and enhances insight into the link between different (allergic) diseases that can coexist in one individual, simultaneously or at different timepoints during a human's life. Endotype-driven thinking can lead to the development of new diagnostic tools, improved therapeutic strategies and better disease management, as well as guide future translational and clinical research into more innovative strategies. These will include new biologics, more effective forms of AIT or even strategies to alter the microbiome and environmental exposures to reduce the risk of allergies. The key discoveries considered in this novel systematology include the role of T-cell subsets, the discovery of the innate mechanisms of non-T2 responses in tissue remodelling and chronicity, the role of virus-induced exacerbations, the epithelial barrier dysfunction, advances in immune metabolism and its consequences on immune polarization contributing to allergic responses. As shortly mentioned above, a combination of different hypersensitivity reactions occurs and develops a mixed type, such as the combination of type I, type IVb, type V and type VI simultaneously. In addition, a conditional skew between different type IV hypersensitivity reactions takes place when there are bacterial, fungal or viral infections over a T2 inflammation in the skin, lung and upper respiratory tissues.

The major advantage of this immune response and tissue-based allergy nomenclature approach is helping move the field towards precision and personalized medicine. The final goal is to tailor the management of individual patients based on their specific immune responses, allergen sensitivities and other factors, including the individual's exposome and metaexposome.<sup>165</sup>

This work will continue to provide advice for clinical practice, which will be conferred in followed-up articles. On top of the

straightforward model shown in this article, the mixed endotypes will be presented and critically discussed because many individuals can express more than one overlapping endotype, which can be a dynamic process over their lifetime.

## AUTHOR CONTRIBUTIONS

Conceptualization: Mark Jutel, Ioana Agache, Cezmi A. Akdis. Writing—original draft preparation: Mark Jutel, Ioana Agache, Cezmi A. Akdis, Magdalena Zemelka-Wiacek, Mübeccel Akdis, Tomás Chivato, Stefano del Giacco, Pawel Gajdanowicz, Ibon Eguluz Gracia, Ludger Klimek, Antti Lauerma, Markus Ollert, Liam O'Mahony, Jürgen Schwarze, Mohamed H. Shamji, Isabel Skypala, Oscar Palomares, Oliver Pfaar, Maria Jose Torres, review and editing: all the authors. All authors have read and agreed to the published version of the manuscript.

## AFFILIATIONS

<sup>1</sup>Department of Clinical Immunology, Wroclaw Medical University, Wroclaw, Poland

<sup>2</sup>ALL-MED Medical Research Institute, Wroclaw, Poland

<sup>3</sup>Faculty of Medicine, Transylvania University, Brasov, Romania

<sup>4</sup>Swiss Institute of Allergy and Asthma Research (SIAF), University of Zurich, Davos, Switzerland

<sup>5</sup>School of Medicine, University CEU San Pablo, Madrid, Spain

<sup>6</sup>Department of Medical Sciences and Public Health, University of Cagliari, Cagliari, Italy

<sup>7</sup>Unit of Allergy and Clinical Immunology, University Hospital "Dulio Casula", Monserrato, Italy

<sup>8</sup>Allergy Unit, UMA-Regional University Hospital of Malaga, IBIMA-BIONAND, Malaga, Spain

<sup>9</sup>Department of Otolaryngology, Head and Neck Surgery, Universitätsmedizin Mainz, Mainz, Germany

<sup>10</sup>Center for Rhinology and Allergy, Wiesbaden, Germany

<sup>11</sup>Department of Dermatology, Helsinki University Hospital and University of Helsinki, Helsinki, Finland

<sup>12</sup>Department of Infection and Immunity, Luxembourg Institute of Health, Esch-sur-Alzette, Luxembourg

<sup>13</sup>Department of Dermatology and Allergy Centre, Odense University Hospital, Odense Research Center for Anaphylaxis (ORCA), Odense, Denmark

<sup>14</sup>Departments of Medicine and Microbiology, APC Microbiome Ireland, National University of Ireland, Cork, Ireland

<sup>15</sup>Child Life and Health, Centre for Inflammation Research, Institute for Regeneration and Repair, The University of Edinburgh, Edinburgh, UK

<sup>16</sup>National Heart and Lung Institute, Imperial College London, London, UK

<sup>17</sup>NIHR Imperial Biomedical Research Centre, London, UK

<sup>18</sup>Department of Inflammation and Repair, Imperial College London, London, UK

<sup>19</sup>Royal Brompton and Harefield Hospitals, Part of Guys and St Thomas' NHS Foundation Trust, London, UK

<sup>20</sup>Department of Biochemistry and Molecular Biology, School of Chemistry, Complutense University of Madrid, Madrid, Spain

<sup>21</sup>Department of Otorhinolaryngology, Head and Neck Surgery, Section of Rhinology and Allergy, University Hospital Marburg, Philipps-Universität Marburg, Marburg, Germany

<sup>22</sup>Department of Internal Medicine, Division of Rheumatology, Allergy and Immunology, University of Cincinnati College of Medicine, Cincinnati, Ohio, USA

<sup>23</sup>Fundação ProAR, Federal University of Bahia and GARD/WHO Planning Group, Salvador, Bahia, Brazil

<sup>24</sup>Allergy and Clinical Immunology, National Heart and Lung Institute, Imperial College London, London, UK

<sup>25</sup>Department of Pathology and Department of Microbiology and Immunology, Stanford University School of Medicine, Stanford, California, USA

<sup>26</sup>Faculty of Health Sciences, Catholic University of Salta, Salta, Argentina

<sup>27</sup>Department of Dermatology and the Laboratory for Inflammatory Skin Diseases, Icahn School of Medicine at Mount Sinai, New York, New York, USA

<sup>28</sup>Skin and Allergy Hospital, Helsinki University Hospital and University of Helsinki, Helsinki, Finland

<sup>29</sup>Academic Unit of Clinical and Experimental Sciences, University of Southampton, Southampton, UK

<sup>30</sup>Department of Biomolecular Sciences, Division of Medical Biochemistry, Saga Medical School, Saga, Japan

<sup>31</sup>Department of Dermatology, Kyoto University Graduate School of Medicine, Kyoto, Japan

<sup>32</sup>Center of Excellence in Asthma and Allergy, Médica Sur Clinical Foundation and Hospital, Mexico City, Mexico

<sup>33</sup>Department of Pediatrics, Dr. von Hauner Children's Hospital, LMU University Hospital, Munich, Germany

<sup>34</sup>Institute of Asthma and Allergy Prevention, Helmholtz Centre Munich, Munich, Germany

<sup>35</sup>German Center for Lung Research (DZL), Giesen, Germany

<sup>36</sup>Department of Environmental Health, Harvard T.H. Chan School of Public Health, Boston, Massachusetts, USA

<sup>37</sup>Department of Pediatrics, Nippon Medical School, Tokyo, Japan

<sup>38</sup>Department of Medicine, Division of Allergy and Clinical Immunology, University of Virginia School of Medicine, Charlottesville, Virginia, USA

<sup>39</sup>Division of Pediatric Allergy and Immunology, Jaffe Food Allergy Institute, Icahn School of Medicine at Mount Sinai, New York, New York, USA

<sup>40</sup>Department of Allergy and Clinical Immunology, Ajou University School of Medicine, Suwon, South Korea

<sup>41</sup>Paul-Ehrlich-Institut, Langen, Germany

<sup>42</sup>Prince of Wales Hospital, Chinese University of Hong Kong, Hong Kong, China

<sup>43</sup>Department of Otolaryngology Head and Neck Surgery, Beijing Tongren Hospital, Capital Medical University, Beijing, China

<sup>44</sup>Beijing Laboratory of Allergic Diseases and Beijing Key Laboratory of Nasal Diseases, Beijing Institute of Otolaryngology, Beijing, China

<sup>45</sup>Department of Clinical and Molecular Sciences, Polytechnic University of Marche, Ancona and Allergy Unit, Department of Internal Medicine, University Hospital of Marche, Ancona, Italy

## ACKNOWLEDGEMENTS

We would like to thank Anna Globinska for the elaboration of all the figures in the Allergy style.

## CONFLICT OF INTEREST STATEMENT

M Jutel reports personal fees from ALK-Abello, Allergopharma, Stallergenes, Anergis, Allergy Therapeutics, Leti, HAL, GSK, Novartis, Teva, Takeda, Chiesi, Pfizer, Regeneron, Astra Zeneca, Lallemand, Shire, CELLTRION Inc. Genetech, Roche, Verona, Lek Pharmaceuticals, Arcutis Biotherapeutics, FAES FARMA outside of submitted work and is the Allergy Journal Deputy Editor and EAACI Board of Officers Member—Past President. I Agache reports being the Allergy Journal Deputy Editor. M Zemelka-Wiacek reports being the EAACI Knowledge Hub Deputy Editor. M Akdis declares grants from Swiss National Science Foundation, European Commission's Horizon's 2020 Framework Programme, Cure. T Chivato reports personal fees from MSD, Uriach and FAES FARMA. He is Vice-president Education and Specialty of EAACI. S del Giacco reports grants from AstraZeneca, GSK, Novartis, Sanofi, outside the submitted work and personal fees from AstraZeneca, Chiesi, CSL-Behring, GSK, Novartis, Sanofi, Stallergenes, Takeda, outside the submitted work; he is the EAACI President. L Klimek reports grants and personal fees from Allergopharma, grants and personal fees



from Viatrix, personal fees from HAL Allergie, personal fees from ALK-Abelló, grants and personal fees from LETI Pharma, grants and personal fees from Stallergenes, grants from Quintiles, grants and personal fees from Sanofi, grants from ASIT biotech, grants from Lofarma, personal fees from Allergy Therapeut., grants from AstraZeneca, grants and personal fees from GSK, grants from Immunotek, personal fees from Cassella med, personal fees from Novartis, personal fees from Regeneron Pharmaceuticals, personal fees from ROXALL Medizin GmbH, outside the submitted work; and Membership: AeDA, DGHNO, Deutsche Akademie für Allergologie und klinische Immunologie, HNO-BV, GPA, EAACI. M Ollert reports personal fees from Hycor Diagnostics, outside the submitted work; and Scientific Co-Founder, Tolerogenics SARL, Luxembourg. L O'Mahony reports personal fees from PrecisionBiotech, grants from GSK and Chiesi, outside the submitted work. He has contributed to company-sponsored symposia for Nestle, Nutricia, Reckitt and Abbott. J Schwarze received consulting fees from Aimune, outside the submitted work. MH Shamji reports grants from Regeneron, Merck, ANGANY Inc, Allergy Therapeutics and Immune Tolerance Network; reports personal fees from Allergopharma; and reports grants and personal fees from ALK, Allergy Therapeutics and ANGANY Inc. I Skypala reports lecture fees from the Royal College of General Practitioners and ThermoFisher, and advisory fees from Touch/ME and is a member of Council of the British Society of Allergy & Clinical Immunology. O Palomares received payment for lectures and participation in Advisory Boards from AstraZeneca, GSK, Pfizer, Immunotek SL, Novartis, Regeneron and Sanofi-Genzyme; received research grants from Novartis SL, Immunotek SL, AstraZeneca, MINECO, MICINNIN and CAM. O Pfaar reports grants and/or personal fees from ALK-Abelló, Allergopharma, Stallergenes Greer, HAL Allergy Holding B.V./HAL Allergie GmbH, Bencard Allergie GmbH/Allergy Therapeutics, Lofarma, ASIT Biotech Tools S.A., Laboratorios LETI/LETI Pharma, GlaxoSmithKline, from ROXALL Medizin, Novartis, Sanofi-Aventis and Sanofi-Genzyme, Med Update Europe GmbH, streamedup! GmbH, Pohl-Boskamp, Immunotek S.L., John Wiley and Sons, AS, Paul-Martini-Stiftung (PMS), Regeneron Pharmaceuticals Inc., RG Aertzfortbildung, Institut für Disease Management, Springer GmbH, AstraZeneca, IQVIA Commercial, Ingress Health, Wort&Bild Verlag, Verlag ME, Procter&Gamble, ALTAMIRA, Meinhardt Congress GmbH, Deutsche Forschungsgemeinschaft, Thieme, Deutsche AllergieLiga e.V., AeDA, Alfried-Krupp Krankenhaus, Red Maple Trials Inc., Königlich Dänisches Generalkonsulat, Medizinische Hochschule Hannover, ECM Expro&Conference Management, Technical University Dresden, all outside the submitted work and within the last 36 months; and he is member of EAACI Excom, member of ext. board of directors DGAKI; coordinator, main- or co-author of different position papers and guidelines in rhinology, allergology and allergen immunotherapy. MJ Torres reports personal fees (report consultancies and speaker bureau) from Diater, Aimune Therapeutics and Leti laboratories, grants from the European Commission, MINECO and ISCIII of the Spanish Government and SEAIC outside the submitted work and is the Allergy Journal

Deputy Editor. JA Bernstein reports receiving consulting fees or payments for research from GSK, AstraZeneca, Sanofi-Regeneron, Amgen, TEVA, ALK, Allergy Therapeutics, Stallergenes, Merck, Genentech and Novartis; is the American Academy of Allergy, Asthma & Immunology (AAAAI) President, a member of the JTF; and is on the board of directors of the World Allergy Organization and Interasma. AA Cruz reports personal fees from AstraZeneca, personal fees from Chiesi, personal fees from GSK, personal fees from SANOFI, personal fees from Novartis, personal fees from Boehringer Ingelheim, personal fees from Glenmark, personal fees from Eurofarma, personal fees from Abdi Ibrahim, personal fees from CROSSJECT, outside the submitted work. SR Durham reports research grants from the Immune Tolerance Network, National Institutes of Allergy and Infectious Diseases USA, Medical Research Council UK and GlaxoSmithKline; has received lecture fees from Abbott Laboratories, ALK, Allergopharma, Pneumo Update GmbH and Stallergenes Greer; and has received consultancy fees from ALK, ANGANY Inc. and Revolo Biotherapeutics. SJ Galli reports grants from the National Institutes of Health (NIH), is a Scientific Advisor of and performs research for, Evommune, Inc. and is a member of the Scientific Advisory Board of Jasper Therapeutics; his patents include 'Multiplex Isotype-Specific Antibody Detection' (U.S. Patent No.: 11,656,233). E Guttman-Yassky Emma Guttman-Yassky is an employee of Mount Sinai and has received research grants from and/or is a consultant for: Research Grants (paid to the institution): Boehringer Ingelheim, Leo Pharma, Pfizer, Cara Therapeutics, UCB, Kyowa Kirin, RAPT, Amgen, GSK, Incyte, Sanofi, Bristol Meyers Squibb, Aslan, Regeneron, Anaptysbio, Concert, Janssen Consultant: Abbvie, Almirall, Amgen, Aslan Pharmaceuticals, AstraZeneca, Biologic Design, Boehringer-Ingelheim, Bristol Meyers Squibb, Cara Therapeutics, Connect Pharma, DBV Technologies, Eli Lilly, EMD Serono, Evidera, Galderma, Gate Bio, Genentech, Incyte, Inmagine, Janssen Biotech, Kyowa Kirin, Leo Pharma, Merck, Pfizer, Q32 Bio, RAPT, Regeneron, Sanofi, SATO, Sioita, Target, UCB, Ventyx. K Izuhara has received grants and personal fees from Shino-test Co. Ltd and grants from AstraZeneca and Maruho. K Kabashima has received consulting fees or advisory board honoraria from Japan Tobacco Inc., Kao, LEO Pharma, Torii, Chugai Pharmaceutical, Maruho, Pola Pharma, Abbvie, Eli Lilly, Sanofi and Pfizer and has received research grants from LEO Pharma, Japan Tobacco Inc., P&G Japan, Eli Lilly Japan, Tanabe Mitsubishi, Ono Pharmaceutical, Kyowa Kirin, Pola Pharma, Abbvie, Sanofi, Kose, Maruho and Kyorin Pharmaceutical. DE Larenas-Linnemann reports personal fees from ALK, AstraZeneca national and global, Bayer, Chiesi, Grunenthal, Grin, GlaxoSmithKline national and global, Viatrix, Karger, Menarini, MSD, Novartis, Pfizer, Sanofi, Siegfried, UCB, Carnot, grants from Abbvie, Bayer, Lilly, Sanofi, AstraZeneca, Pfizer, Novartis, Circassia, UCB, GlaxoSmithKline, outside the submitted work. E von Mutius reports grants from the German Federal Ministry of Education and Research (BMBF), German Center for Lung Research, Bavarian State Ministry of Health and Care for 'URS Study', 01/21, 'Impact Chip Study', O.M. Pharma S.A. She reports Royalties or licenses from Elsevier GmbH, Georg Thieme

Verlag, Springer-Verlag GmbH, Elsevier Ltd., Springer Nature Group. She reports payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events Massachusetts Medical Society, Springer-Verlag GmbH, Elsevier Ltd., Böhringer Ingelheim International GmbH, European Respiratory Society (ERS), Universiteit Utrecht, Faculteit Diergeneeskunde, Universität Salzburg, Springer Medizin Verlag GmbH, Japanese Society of Pediatric Allergy and Clinical Immunology (JSPACI), Klinikum Rechts der Isar, University of Colorado, Paul-Martini-Stiftung, Astra Zeneca, Imperial College London, Children's Hospital Research Institute of Manitoba, Kompetenzzentrum für Ernährung (Kern), OM Pharma S.A., Swedish Pediatric Society for Allergy and Lung Medicine, Chinese College of Allergy and Asthma (CCAA), ALK-Abello Arzneimittel GmbH, Abbott Laboratories, Deutscher Apotheker Verlag GmbH & Co. KG, Japanese Society of Allergology, British Society for Asthma and Clinical Immunology, American Academy of Allergy, Asthma & Immunology, OM Pharma S.A. She reports support for attending meetings from Verein zur Förderung der Pneumologie am Krankenhaus Großhansdorf e.V., Pneumologie Developpement, Mondial Congress & Events GmbH & Co. KG, American Academy of Allergy, Asthma & Immunology, Imperial College London, Margaux Orange, Volkswagen Stiftung, Böhringer Ingelheim International GmbH, European Respiratory Society (ERS), Universiteit Utrecht, Faculteit Diergeneeskunde, Österreichische Gesellschaft f. Allergologie u. Immunologie, Massachusetts Medical Society, OM Pharma S. A., Hanson Wade Ltd., iKOMM GmbH, DSI Dansk Borneastma Center, American Thoracic Society, HiPP GmbH & Co KG, Universiteit Utrecht, Faculteit Bètawetenschappen, ALK-Abello Arzneimittel GmbH, Deutsches Zentrum für Lungenforschung (DZL), Fabio Luigi Massimo Ricciardolo/Contatto S.r.l., Fraunhofer ITEM Hannover, MCCA Institut für Immunologie Uni Wien, Swiss Institute of Allergy and Asthma Research (SIAF) Davos (Associated Institute of the University of Zurich), MHH (Medizinische Hochschule Hannover, European Respiratory Society, Natasha Allergy Research Foundation, Deutsche Forschungsgemeinschaft, Gordon Research Conferences, Societed Chilena de Enfermedades Respiratorias, Arla, Universität Leiden, American Academy of Allergy, Asthma & Clinical Immunology, Deutsche Forschungsgemeinschaft (DFG), ERS, Deutsches Zentrum für Lungenforschung. She reports patents planned, issued or pending from EvM has patent No. PCT/EP2019/085016 (Barn dust extract for the prevention and treatment of diseases) pending (Barn dust extract for the prevention and treatment of diseases) pending, royalties paid to ProtectImmun for patent EP2361632 (Specific environmental bacteria for the protection from and/or the treatment of allergic, chronic inflammatory and/or autoimmune disorders, granted on 19 March 2014) and patents EP1411977 (Composition containing bacterial antigens used for the prophylaxis and the treatment of allergic diseases, granted on 18 April 2007), EP1637147 (Stable dust extract for allergy protection, granted on 10 December 2008) and EP 1964570 (Pharmaceutical compound to protect against allergies and inflammatory diseases, granted on 21 November 2012) licensed to ProtectImmun. Patent

EP21189353.2. 2021. von Mutius E, Rankl B, Bracher F, Müller C, Walker A, Hauck SM, Merl-Pham J, inventors; Proteins identified from barn dust extract for the prevention and treatment of diseases. Patent PCT/US2021/016918. 2021. Martinez FD, Vercelli D, Snyder SA, von Mutius E, Pivniouk V, Marques dos Santos M, inventors; Therapeutic fractions and proteins from asthma-protective farm dust. Patent EP21189353.2. 2021. von Mutius E, Rankl B, Bracher F, Müller C, Walker A, Hauck SM, Merl-Pham J, Adler H, Yildirim A.Ö., Sattler M, Santos Dias Mourao A, Borggräfe J, O'Connor P.D., Plettenburg O, inventors; Proteins identified from barn dust extract for the prevention and treatment of diseases. She report participation on a Data Safety Monitoring Board or Advisory Board from Member of the EXPANSE (funded by European Commission) Scientific Advisory Board, Member of the BEAMS External Scientific Advisory Board (ESAB), Member of the Editorial Board of 'The Journal of Allergy and Clinical Immunology: In Practice', Member of the Scientific Advisory Board of the Children's Respiratory and Environmental Workgroup (CREW), Member of the International Scientific & Societal Advisory Board (ISSAB) of Utrecht Life Sciences (ULS), University of Utrecht, Member of External Review Panel of the Faculty of Veterinary Science, University of Utrecht, Member of the Selection Committee for the Gottfried Wilhelm Leibniz Programme (DFG), Member of the International Advisory Board of Asthma UK Centre for Applied Research (AUKCAR), Member of the International Advisory Board of 'The Lancet Respiratory Medicine', Member of the Scientific Advisory Board of the CHILD (Canadian Healthy Infant Longitudinal Development) study, McMaster University, Hamilton, Canada, Asthma UK Centre for Applied Research, Pediatric Scientific Advisory Board Iceland, Abbott Allergy Risk Reduction Advisory Board. KC Nadeau reports grants from the National Institute of Allergy and Infectious Diseases (NIAID), National Heart, Lung and Blood Institute (NHLBI), National Institute of Environmental Health Sciences (NIEHS) and Food Allergy Research & Education (FARE); Stock options from IgGenix, Seed Health, ClostraBio, Cour, Alladapt, Clostrabio and ImmunelD; Director of the World Allergy Organization Center of Excellence for Stanford; Advisor at Cour Pharma; Consultant for Excellergy, Red tree ventures, Before Brands, Alladapt, Cour, Latitude, Regeneron and IgGenix; Co-founder of Before Brands, Alladapt, Latitude and IgGenix; National Scientific Committee member at Immune Tolerance Network (ITN) and National Institutes of Health (NIH) clinical research centers; patents include, 'Mixed allergen com-position and methods for using the same', 'Granulocyte-based methods for detecting and monitoring immune system disorders', and 'Methods and Assays for Detecting and Quantifying Pure Subpopulations of White Blood Cells in Immune System Disorders'. SH Sicherer reports royalty payments from UpToDate and from Johns Hopkins University Press; grants to his institution from the National Institute of Allergy and Infectious Diseases, from Food Allergy Research and Education and Pfizer; and personal fees from the American Academy of Allergy, Asthma and Immunology as Deputy Editor of the Journal of Allergy and Clinical Immunology: In Practice, outside of the submitted work. HS Park reports

research grants from Sanofi-Aventis and YH and clinical trial funds from Astra-Zeneca and Teva-Korea. S Vieths reports personal fees from Schattauer Allergologie Handbuch, Elsevier Nahrungsmittelallergien und Intoleranzen, Karger Food Allergy: Molecular Basis and Clinical Practice and as Associate Editor of the Journal of Allergy and Clinical Immunology, all outside the submitted work. MB Bilò reports personal fees from AlkAbello, AstraZeneca, Chiesi, GSK, Novartis, Sanofi, outside the submitted work. CA Akdis has received research grants from the Swiss National Science Foundation, European Union (EU CURE, EU Syn-Air-G), Novartis Research Institutes, (Basel, Switzerland), Stanford University (Redwood City, Calif) and SciBase (Stockholm, Sweden); is the Co-Chair for EAACI Guidelines on Environmental Science in Allergic diseases and Asthma; is on the Advisory Boards of Sanofi/Regeneron (Bern, Switzerland, New York, USA), Stanford University Sean Parker Asthma Allergy Center (CA, USA), Novartis (Basel, Switzerland), Glaxo Smith Kline (Zurich, Switzerland), Bristol-Myers Squibb (New York, USA), Seed Health (Boston, USA) and SciBase (Stockholm, Sweden); and is the Editor-in-Chief of Allergy Journal. P Gajdanowicz, IE Gracia, A Lauerma, RM Gómez, T Haahtela, ST Holgate, R Pawankar, TAE Platts-Mills, G Wong and L Zhang have nothing to disclose.

#### DATA AVAILABILITY STATEMENT

Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

#### ORCID

Marek Jutel  <https://orcid.org/0000-0003-1555-9379>  
 Ioana Agache  <https://orcid.org/0000-0001-7994-364X>  
 Magdalena Zemelka-Wiacek  <https://orcid.org/0000-0001-7201-8638>  
 Mübeccel Akdis  <https://orcid.org/0000-0003-0554-9943>  
 Tomás Chivato  <https://orcid.org/0000-0002-5403-0964>  
 Stefano del Giacco  <https://orcid.org/0000-0002-4517-1749>  
 Pawel Gajdanowicz  <https://orcid.org/0000-0003-3075-5992>  
 Ibon Eguiluz Gracia  <https://orcid.org/0000-0002-3774-931X>  
 Ludger Klimek  <https://orcid.org/0000-0002-2455-0192>  
 Antti Lauerma  <https://orcid.org/0000-0002-5078-3547>  
 Markus Ollert  <https://orcid.org/0000-0002-8055-0103>  
 Liam O'Mahony  <https://orcid.org/0000-0003-4705-3583>  
 Jurgen Schwarze  <https://orcid.org/0000-0002-6899-748X>  
 Mohamed H. Shamji  <https://orcid.org/0000-0003-3425-3463>  
 Isabel Skypala  <https://orcid.org/0000-0003-3629-4293>  
 Oscar Palomares  <https://orcid.org/0000-0003-4516-0369>  
 Oliver Pfaar  <https://orcid.org/0000-0003-4374-9639>  
 Maria Jose Torres  <https://orcid.org/0000-0001-5228-471X>  
 Jonathan A. Bernstein  <https://orcid.org/0000-0002-3476-1196>  
 Alvaro A. Cruz  <https://orcid.org/0000-0002-7403-3871>  
 Stephen R. Durham  <https://orcid.org/0000-0001-5264-6207>  
 Stephen J. Galli  <https://orcid.org/0000-0001-5736-5340>  
 R. Maximiliano Gomez  <https://orcid.org/0000-0001-6898-186X>

Emma Guttman-Yassky  <https://orcid.org/0000-0002-9363-324X>  
 Tari Haahtela  <https://orcid.org/0000-0003-4757-2156>  
 Stephen T. Holgate  <https://orcid.org/0000-0003-2658-4617>  
 Kenji Izuhara  <https://orcid.org/0000-0002-6983-907X>  
 Kenji Kabashima  <https://orcid.org/0000-0002-0773-0554>  
 Désirée E. Larenas-Linnemann  <https://orcid.org/0000-0002-5713-5331>  
 Erica von Mutius  <https://orcid.org/0000-0002-8893-4515>  
 Kari C. Nadeau  <https://orcid.org/0000-0002-2146-2955>  
 Ruby Pawankar  <https://orcid.org/0000-0002-3091-7237>  
 Tomas A. E. Platts-Mills  <https://orcid.org/0000-0002-1263-329X>  
 Scott H. Sicherer  <https://orcid.org/0000-0003-0036-0439>  
 Hae-Sim Park  <https://orcid.org/0000-0003-2614-0303>  
 Stefan Vieths  <https://orcid.org/0000-0003-3826-6704>  
 Gary Wong  <https://orcid.org/0000-0001-5939-812X>  
 Luo Zhang  <https://orcid.org/0000-0002-0910-9884>  
 M. Beatrice Bilò  <https://orcid.org/0000-0002-9324-6039>  
 Cezmi A. Akdis  <https://orcid.org/0000-0001-8020-019X>

#### REFERENCES

- Lötvall J, Akdis CA, Bacharier LB, et al. Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome. *J Allergy Clin Immunol.* 2011;127(2):355-360. doi:10.1016/j.jaci.2010.11.037
- Ozdemir C, Kucuksezer UC, Akdis M, Akdis CA. The concepts of asthma endotypes and phenotypes to guide current and novel treatment strategies. *Expert Rev Respir Med.* 2018;12(9):733-743. doi:10.1080/17476348.2018.1505507
- Agache I, Akdis CA. Endotypes of allergic diseases and asthma: an important step in building blocks for the future of precision medicine. *Allergol Int.* 2016;65(3):243-252. doi:10.1016/j.alit.2016.04.011
- Agache I, Eguiluz-Gracia I, Cojanu C, et al. Advances and highlights in asthma in 2021. *Allergy.* 2021;76(11):3390-3407. doi:10.1111/all.15054
- Papadopoulos NG, Bernstein JA, Demoly P, et al. Phenotypes and endotypes of rhinitis and their impact on management: a PRACTALL report. *Allergy.* 2015;70(5):474-494. doi:10.1111/all.12573
- Papadopoulos NG, Guibas GV. Rhinitis subtypes, endotypes, and definitions. *Immunol Allergy Clin North Am.* 2016;36(2):215-233. doi:10.1016/j.iac.2015.12.001
- Segboer CL, Fokkens WJ, Terreehorst I, van Drunen CM. Endotyping of non-allergic, allergic and mixed rhinitis patients using a broad panel of biomarkers in nasal secretions. *PLoS One.* 2018;13(7):e0200366. doi:10.1371/journal.pone.0200366
- Akdis CA, Bachert C, Cingi C, et al. Endotypes and phenotypes of chronic rhinosinusitis: a PRACTALL document of the European academy of allergy and clinical immunology and the American Academy of Allergy, Asthma & Immunology. *J Allergy Clin Immunol.* 2013;131(6):1479-1490. doi:10.1016/j.jaci.2013.02.036
- Kato A, Peters AT, Stevens WW, Schleimer RP, Tan BK, Kern RC. Endotypes of chronic rhinosinusitis: relationships to disease phenotypes, pathogenesis, clinical findings, and treatment approaches. *Allergy.* 2022;77(3):812-826. doi:10.1111/all.15074
- Thijs JL, Strickland I, Bruijnzeel-Koomen CAFM, et al. Moving toward endotypes in atopic dermatitis: identification of patient clusters based on serum biomarker analysis. *J Allergy Clin Immunol.* 2017;140(3):730-737. doi:10.1016/j.jaci.2017.03.023

11. Czarnewicki T, He H, Krueger JG, Guttman-Yassky E. Atopic dermatitis endotypes and implications for targeted therapeutics. *J Allergy Clin Immunol*. 2019;143(1):1-11. doi:10.1016/j.jaci.2018.10.032
12. Hui-Beckman JW, Goleva E, Berdyshev E, Leung DYM. Endotypes of atopic dermatitis and food allergy. *J Allergy Clin Immunol*. 2023;151(1):26-28. doi:10.1016/j.jaci.2022.07.021
13. Blank S, Grosch J, Ollert M, Bilò MB. Precision medicine in hymenoptera venom allergy: diagnostics, biomarkers, and therapy of different Endotypes and phenotypes. *Front Immunol*. 2020;22(11):579409. doi:10.3389/fimmu.2020.579409
14. Muraro A, Lemanske RF Jr, Castells M, et al. Precision medicine in allergic disease-food allergy, drug allergy, and anaphylaxis-PRAC-TALL document of the European academy of allergy and clinical immunology and the American Academy of Allergy, Asthma and Immunology. *Allergy*. 2017;72(7):1006-1021. doi:10.1111/all.13132
15. Agache I, Shamji MH, Kermani NZ, et al. Multidimensional endotyping using nasal proteomics predicts molecular phenotypes in the asthmatic airways. *J Allergy Clin Immunol*. 2023;151(1):128-137. doi:10.1016/j.jaci.2022.06.028
16. Agache I, Akdis CA. Precision medicine and phenotypes, endotypes, genotypes, regiotypes, and theratypes of allergic diseases. *J Clin Invest*. 2019;129(4):1493-1503. doi:10.1172/JCI124611
17. Agache I, Akdis C, Jutel M, Virchow JC. Untangling asthma phenotypes and endotypes. *Allergy*. 2012;67(7):835-846. doi:10.1111/j.1398-9995.2012.02832.x
18. Dyjack N, Goleva E, Rios C, et al. Minimally invasive skin tape strip RNA sequencing identifies novel characteristics of the type 2-high atopic dermatitis disease endotype. *J Allergy Clin Immunol*. 2018;141(4):1298-1309. doi:10.1016/j.jaci.2017.10.046
19. Shamji MH, Ollert M, Adcock IM, et al. EAACI guidelines on environmental science in allergic diseases and asthma-leveraging artificial intelligence and machine learning to develop a causality model in exposomics. *Allergy*. 2023;78:1742-1757. doi:10.1111/all.15667
20. Bendiner E. Baron von Pirquet: the aristocrat who discovered and defined allergy. *Hosp Pract (off Ed)*. 1981;16(10):137-141.
21. Johansson SG, Hourihane JO, Bousquet J, et al. A revised nomenclature for allergy. An EAACI position statement from the EAACI nomenclature task force. *Allergy*. 2001;56(9):813-824. doi:10.1034/j.1398-9995.2001.t01-1-00001.x
22. Gell PG. Studies of undernutrition Wuppertal 1946-9. XI. Serological responses to antigenic stimuli. *Spec Rep Ser Med Res Counc (G B)*. 1951;275:193-203.
23. Gell PG, Hinde IT. The histology of the tuberculin reaction and its modification by cortisone. *Br J Exp Pathol*. 1951;32(6):516-529.
24. Gell PG, Hinde IT. The effect of cortisone on the histology of the tuberculin reaction. *Bull Schweiz Akad Med Wiss*. 1952;8(1-2):200-202.
25. Coombs PR, Gell PG. Classification of allergic reactions responsible for clinical hypersensitivity and disease. In: Gell RR, ed. *Clinical Aspects of Immunology*. Oxford University Press; 1968:575-596.
26. Johansson SG, Bieber T, Dahl R, et al. Revised nomenclature for allergy for global use: report of the nomenclature review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol*. 2004;113(5):832-836. doi:10.1016/j.jaci.2003.12.591
27. Del Prete GF, De Carli M, Mastromauro C, et al. Purified protein derivative of *Mycobacterium tuberculosis* and excretory-secretory antigen(s) of *Toxocara canis* expand in vitro human T cells with stable and opposite (type 1 T helper or type 2 T helper) profile of cytokine production. *J Clin Invest*. 1991;88(1):346-350. doi:10.1172/JCI115300
28. Wierenga EA, Snoek M, Jansen HM, Bos JD, van Lier RA, Kapsenberg ML. Human atopen-specific types 1 and 2 T helper cell clones. *J Immunol*. 1991;147(9):2942-2949.
29. Robinson DS, Hamid Q, Ying S, et al. Predominant TH2-like bronchoalveolar T-lymphocyte population in atopic asthma. *N Engl J Med*. 1992;326(5):298-304. doi:10.1056/NEJM199201303260504
30. Pichler WJ. Delayed drug hypersensitivity reactions. *Ann Intern Med*. 2003;139(8):683-693. doi:10.7326/0003-4819-139-8-200310210-00012
31. Posadas SJ, Pichler WJ. Delayed drug hypersensitivity reactions—new concepts. *Clin Exp Allergy*. 2007;37(7):989-999. doi:10.1111/j.1365-2222.2007.02742.x
32. Hausmann O, Schnyder B, Pichler WJ. Drug hypersensitivity reactions involving skin. *Handb Exp Pharmacol*. 2010;196:29-55. doi:10.1007/978-3-642-00663-0\_2
33. Akdis CA, Arkwright PD, Brügggen MC, et al. Type 2 immunity in the skin and lungs. *Allergy*. 2020;75(7):1582-1605. doi:10.1111/all.14318
34. Annunziato F, Romagnani C, Romagnani S. The 3 major types of innate and adaptive cell-mediated effector immunity. *J Allergy Clin Immunol*. 2015;135(3):626-635. doi:10.1016/j.jaci.2014.11.001
35. Han X, Krempski JW, Nadeau K. Advances and novel developments in mechanisms of allergic inflammation. *Allergy*. 2020;75(12):3100-3111. doi:10.1111/all.14632
36. Zheng H, Zhang Y, Pan J, et al. The role of type 2 innate lymphoid cells in allergic diseases. *Front Immunol*. 2021;12:586078. doi:10.3389/fimmu.2021.586078
37. Agache I, Zemelka-Wiącek M, Shamji MH, Jutel M. Immunotherapy: state-of-the-art review of therapies and theratypes. *J Allergy Clin Immunol*. 2022;150(6):1279-1288. doi:10.1016/j.jaci.2022.10.007
38. Simons FE, Arduoso LR, Bilò MB, et al. World allergy organization guidelines for the assessment and management of anaphylaxis. *World Allergy Organ J*. 2011;4(2):13-37. doi:10.1097/WOX.0b013e318211496c
39. Cardona V, Ansotegui IJ, Ebisawa M, et al. World allergy organization anaphylaxis guidance 2020. *World Allergy Organ J*. 2020;13(10):100472. doi:10.1016/j.waojou.2020.100472
40. Justiz Vaillant AA, Vashisht R, Zito PM. Immediate hypersensitivity reactions. *StatPearls [Internet]*. StatPearls Publishing; 2022.
41. Balan S, Saxena M, Bhardwaj N. Dendritic cell subsets and locations. *Int Rev Cell Mol Biol*. 2019;348:1-68. doi:10.1016/bs.ircmb.2019.07.004
42. Gauvreau GM, Bergeron C, Boulet LP, et al. Sounding the alarms—the role of alarmin cytokines in asthma. *Allergy*. 2023;78(2):402-417. doi:10.1111/all.15609
43. Saito K, Orimo K, Kubo T, et al. Laundry detergents and surfactants induced eosinophilic airway inflammation by increasing IL-33 expression and activating ILC2s. *Allergy*. 2023;78:1878-1892. doi:10.1111/all.15762
44. Möller KJ, Wegner L, Malsy J, et al. Expanded ILC2s in human infant intestines promote tissue-growth. *Mucosal Immunol*. 2023;16(4):408-421. doi:10.1016/j.mucimm.2023.04.004
45. Pelly VS, Kannan Y, Coomes SM, et al. IL-4-producing ILC2s are required for the differentiation of TH2 cells following *Heligmosomoides polygyrus* infection. *Mucosal Immunol*. 2016;9(6):1407-1417. doi:10.1038/mi.2016.4
46. Jin J, Sunusi S, Lu H. Group 2 innate lymphoid cells (ILC2s) are important in typical type 2 immune-mediated diseases and an essential therapeutic target. *J Int Med Res*. 2022;50(1):3000605211053156. doi:10.1177/03000605211053156
47. Varricchi G, Bencivenga L, Poto R, Pecoraro A, Shamji MH, Rengo G. The emerging role of T follicular helper (TFH) cells in aging: influence on the immune frailty. *Ageing Res Rev*. 2020;61:101071. doi:10.1016/j.arr.2020.101071

48. Akdis CA, Blesken T, Akdis M, Wüthrich B, Blaser K. Role of interleukin 10 in specific immunotherapy. *J Clin Invest*. 1998;102(1):98-106. doi:10.1172/JCI2250
49. Jutel M, Akdis M, Budak F, et al. IL-10 and TGF-beta cooperate in the regulatory T cell response to mucosal allergens in normal immunity and specific immunotherapy. *Eur J Immunol*. 2003;33(5):1205-1214. doi:10.1002/eji.200322919
50. Nouri-Aria KT, Wachholz PA, Francis JN, et al. Grass pollen immunotherapy induces mucosal and peripheral IL-10 responses and blocking IgG activity. *J Immunol*. 2004;172(5):3252-3259. doi:10.4049/jimmunol.172.5.3252
51. van de Veen W, Stanic B, Yaman G, et al. IgG4 production is confined to human IL-10-producing regulatory B cells that suppress antigen-specific immune responses. *J Allergy Clin Immunol*. 2013;131(4):1204-1212. doi:10.1016/j.jaci.2013.01.014
52. Morita H, Kubo T, Rückert B, et al. Induction of human regulatory innate lymphoid cells from group 2 innate lymphoid cells by retinoic acid. *J Allergy Clin Immunol*. 2019;143(6):2190-2201.e9. doi:10.1016/j.jaci.2018.12.1018
53. Varricchi G, Harker J, Borriello F, Marone G, Durham SR, Shamji MH. T follicular helper (Tfh) cells in normal immune responses and in allergic disorders. *Allergy*. 2016;71(8):1086-1094. doi:10.1111/all.12878
54. Moon TC, Befus AD, Kulka M. Mast cell mediators: their differential release and the secretory pathways involved. *Front Immunol*. 2014;5:569. doi:10.3389/fimmu.2014.00569
55. Stone KD, Prussin C, Metcalfe DD. IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol*. 2010;125(2 Suppl 2):S73-S80. doi:10.1016/j.jaci.2009.11.017
56. Bellinghausen I, Khatri R, Saloga J. Current strategies to modulate regulatory T cell activity in allergic inflammation. *Front Immunol*. 2022;13:912529. doi:10.3389/fimmu.2022.912529
57. Qin L, Tang LF, Cheng L, Wang HY. The clinical significance of allergen-specific IgG4 in allergic diseases. *Front Immunol*. 2022;13:1032909. doi:10.3389/fimmu.2022.1032909
58. Pilette C, Nouri-Aria KT, Jacobson MR, et al. Grass pollen immunotherapy induces an allergen-specific IgA2 antibody response associated with mucosal TGF-beta expression. *J Immunol*. 2007;178(7):4658-4666. doi:10.4049/jimmunol.178.7.4658
59. Gigon L, Fettlelet T, Yousefi S, Simon D, Simon HU. Eosinophils from a to Z. *Allergy*. 2023;78(7):1810-1846. doi:10.1111/all.15751
60. Lee E, Kim M, Jeon K, et al. Mean platelet volume, platelet distribution width, and platelet count, in connection with immune thrombocytopenic purpura and essential thrombocytopenia. *Lab Med*. 2019;50(3):279-285. doi:10.1093/labmed/lmy082
61. Li TX, Sun FT, Ji BJ. Correlation of IgG subclass with blood cell parameters in patients with autoimmune hemolytic anemia. *Zhongguo Shi Yan Xue Ye Xue Za Zhi*. 2019;27(1):197-201. doi:10.7534/j.issn.1009-2137.2019.01.032
62. Leonard A, Hittson Boal L, Pary P, et al. Identification of red blood cell antibodies in maternal breast milk implicated in prolonged hemolytic disease of the fetus and newborn. *Transfusion*. 2019;59(4):1183-1189. doi:10.1111/trf.15154
63. Vries TB, Boerma S, Doornebal J, Dikkeschei B, Stegeman C, Veneman TF. Goodpasture's syndrome with negative anti-glomerular basement membrane antibodies. *Eur J Case Rep Intern Med*. 2017;4(8):000687. doi:10.12890/2017\_000687
64. Wang W, Erbe AK, Hank JA, Morris ZS, Sondel PM. NK cell-mediated antibody-dependent cellular cytotoxicity in cancer immunotherapy. *Front Immunol*. 2015;6:368. doi:10.3389/fimmu.2015.00368
65. Sauler M, Bazan IS, Lee PJ. Cell death in the lung: the apoptosis-necroptosis Axis. *Annu Rev Physiol*. 2019;81:375-402. doi:10.1146/annurev-physiol-020518-114320
66. Dai W, Cheng J, Leng X, Hu X, Ao Y. The potential role of necroptosis in clinical diseases (review). *Int J Mol Med*. 2021;47(5):89. doi:10.3892/ijmm.2021.4922
67. Bajwa SF, Mohammed RH. Type II hypersensitivity reaction. *StatPearls [Internet]*. StatPearls Publishing; 2023.
68. Usman N, Annamaraju P. Type III hypersensitivity reaction. *StatPearls [Internet]*. StatPearls Publishing; 2022.
69. Ramos BF, Zhang Y, Jakschik BA. Neutrophil elicitation in the reverse passive Arthus reaction. Complement-dependent and -independent mast cell involvement. *J Immunol*. 1994;152(3):1380-1384.
70. Sylvestre DL, Ravetch JV. A dominant role for mast cell fc receptors in the Arthus reaction. *Immunity*. 1996;5(4):387-390. doi:10.1016/s1074-7613(00)80264-2
71. Sylvestre DL, Ravetch JV. Fc receptors initiate the Arthus reaction: redefining the inflammatory cascade. *Science*. 1994;265(5175):1095-1098. doi:10.1126/science.8066448
72. Krystel-Whittemore M, Dileepan KN, Wood JG. Mast cell: a multi-functional master cell. *Front Immunol*. 2016;6:620. doi:10.3389/fimmu.2015.00620
73. Shushakova N, Skokowa J, Schulman J, et al. C5a anaphylatoxin is a major regulator of activating versus inhibitory FcγR3s in immune complex-induced lung disease. *J Clin Invest*. 2002;110(12):1823-1830. doi:10.1172/JCI16577
74. Romano A, Valluzzi RL, Caruso C, Maggioletti M, Gaeta F. Non-immediate cutaneous reactions to Beta-lactams: approach to diagnosis. *Curr Allergy Asthma Rep*. 2017;17(4):23. doi:10.1007/s11882-017-0691-4
75. Pan J, Zhang M, Wang J, et al. Interferon-gamma is an autocrine mediator for dendritic cell maturation. *Immunol Lett*. 2004;94(1-2):141-151. doi:10.1016/j.imlet.2004.05.003
76. McLaughlin TA, Khayumbi J, Ongalo J, et al. CD4 T cells in *Mycobacterium tuberculosis* and *Schistosoma mansoni* Co-infected individuals maintain functional TH1 responses. *Front Immunol*. 2020;11:127. doi:10.3389/fimmu.2020.00127
77. Seillet C, Belz GT, Huntington ND. Development, homeostasis, and heterogeneity of NK cells and ILC1. *Curr Top Microbiol Immunol*. 2016;395:37-61. doi:10.1007/82\_2015\_474
78. Trautmann A, Akdis M, Kleemann D, et al. T cell-mediated Fas-induced keratinocyte apoptosis plays a key pathogenetic role in eczematous dermatitis. *J Clin Invest*. 2000;106(1):25-35. doi:10.1172/JCI9199
79. Trautmann A, Schmid-Grendelmeier P, Krüger K, et al. T cells and eosinophils cooperate in the induction of bronchial epithelial cell apoptosis in asthma. *J Allergy Clin Immunol*. 2002;109(2):329-337. doi:10.1067/mai.2002.121460
80. Preglej T, Hamminger P, Luu M, et al. Histone deacetylases 1 and 2 restrain CD4+ cytotoxic T lymphocyte differentiation. *JCI Insight*. 2020;5(4):e133393. doi:10.1172/jci.insight.133393
81. Bhat P, Leggatt G, Waterhouse N, Frazer IH. Interferon-γ derived from cytotoxic lymphocytes directly enhances their motility and cytotoxicity. *Cell Death Dis*. 2017;8(6):e2836. doi:10.1038/cddis.2017.67
82. Valkenburg SA, Gras S, Guilloncneau C, et al. Protective efficacy of cross-reactive CD8+ T cells recognising mutant viral epitopes depends on peptide-MHC-I structural interactions and T cell activation threshold. *PLoS Pathog*. 2010;6(8):e1001039. doi:10.1371/journal.ppat.1001039
83. Zimmermann M, Koreck A, Meyer N, et al. TNF-like weak inducer of apoptosis (TWEAK) and TNF-α cooperate in the induction of keratinocyte apoptosis. *J Allergy Clin Immunol*. 2011;127(1):200-207. doi:10.1016/j.jaci.2010.11.005
84. Rebane A, Zimmermann M, Aab A, et al. Mechanisms of IFN-γ induced apoptosis of human skin keratinocytes in patients with atopic dermatitis. *J Allergy Clin Immunol*. 2012;129(5):1297-1306. doi:10.1016/j.jaci.2012.02.020

85. Valente M, Dölen Y, van Dinther E, et al. Cross-talk between iNKT cells and CD8 T cells in the spleen requires the IL-4/CCL17 axis for the generation of short-lived effector cells. *Proc Natl Acad Sci U S A*. 2019;116(51):25816-25827. doi:10.1073/pnas.1913491116
86. Eguiluz-Gracia I, Layhadi JA, Rondon C, Shamji MH. Mucosal IgE immune responses in respiratory diseases. *A-Curr Opin Pharmacol*. 2019;46:100-107. doi:10.1016/j.coph.2019.05.009
87. Fahy JV. Type 2 inflammation in asthma—Present in most, absent in many. *Nat Rev Immunol*. 2015;15(1):57-65. doi:10.1038/nri3786
88. Datsi A, Steinhoff M, Ahmad F, Alam M, Buddenkotte J. Interleukin-31: the “itchy” cytokine in inflammation and therapy. *Allergy*. 2021;76(10):2982-2997. doi:10.1111/all.14791
89. Garcovich S, Maurelli M, Gisoni P, Peris K, Yosipovitch G, Girolomoni G. Pruritus as a distinctive feature of type 2 inflammation. *Vaccines (Basel)*. 2021;9(3):303. doi:10.3390/vaccines9030303
90. Abdelaziz MH, Wang H, Cheng J, Xu H. Th2 cells as an intermediate for the differentiation of naïve T cells into Th9 cells, associated with the Smad3/Smad4 and IRF4 pathway. *Exp Ther Med*. 2020;19(3):1947-1954. doi:10.3892/etm.2020.8420
91. Kaplan MH, Hufford MM, Olson MR. The development and in vivo function of T helper 9 cells. *Nat Rev Immunol*. 2015;15(5):295-307. doi:10.1038/nri3824
92. Starkey MR, McKenzie AN, Belz GT, Hansbro PM. Pulmonary group 2 innate lymphoid cells: surprises and challenges. *Mucosal Immunol*. 2019;12(2):299-311. doi:10.1038/s41385-018-0130-4
93. Orimo K, Tamari M, Saito H, Matsumoto K, Nakae S, Morita H. Characteristics of tissue-resident ILCs and their potential as therapeutic targets in mucosal and skin inflammatory diseases. *Allergy*. 2021;76(11):3332-3348. doi:10.1111/all.14863
94. Wawrzyniak P, Wawrzyniak M, Wanke K, et al. Regulation of bronchial epithelial barrier integrity by type 2 cytokines and histone deacetylases in asthmatic patients. *J Allergy Clin Immunol*. 2017;139(1):93-103. doi:10.1016/j.jaci.2016.03.050
95. Sugita K, Steer CA, Martinez-Gonzalez I, et al. Type 2 innate lymphoid cells disrupt bronchial epithelial barrier integrity by targeting tight junctions through IL-13 in asthmatic patients. *J Allergy Clin Immunol*. 2018;141(1):300-310.e11. doi:10.1016/j.jaci.2017.02.038
96. Zhu J. T helper 2 (Th2) cell differentiation, type 2 innate lymphoid cell (ILC2) development and regulation of interleukin-4 (IL-4) and IL-13 production. *Cytokine*. 2015;75(1):14-24. doi:10.1016/j.cyto.2015.05.010
97. Miyake K, Shibata S, Yoshikawa S, Karasuyama H. Basophils and their effector molecules in allergic disorders. *Allergy*. 2021;76(6):1693-1706. doi:10.1111/all.14662
98. Akdis M, Aab A, Altunbulakli C, et al. Interleukins (from IL-1 to IL-38), interferons, transforming growth factor  $\beta$ , and TNF- $\alpha$ : receptors, functions, and roles in diseases. *J Allergy Clin Immunol*. 2016;138(4):984-1010. doi:10.1016/j.jaci.2016.06.033
99. Hancock A, Armstrong L, Gama R, Millar A. Production of interleukin 13 by alveolar macrophages from normal and fibrotic lung. *Am J Respir Cell Mol Biol*. 1998;18(1):60-65. doi:10.1165/ajrcmb.18.1.2627
100. Bochner BS. Systemic activation of basophils and eosinophils: markers and consequences. *J Allergy Clin Immunol*. 2000;106(5 Suppl):S292-S302. doi:10.1067/mai.2000.110164
101. Valent P, Klion AD, Roufosse F, et al. Proposed refined diagnostic criteria and classification of eosinophil disorders and related syndromes. *Allergy*. 2023;78(1):47-59. doi:10.1111/all.15544
102. Thompson-Souza GA, Vasconcelos CRI, Neves JS. Eosinophils: focus on DNA extracellular traps. *Life Sci*. 2022;311(Pt B):121191. doi:10.1016/j.lfs.2022.121191
103. Johnson MO, Wolf MM, Madden MZ, et al. Distinct regulation of Th17 and Th1 cell differentiation by Glutaminase-dependent metabolism. *Cell*. 2018;175(7):1780-1795.e19. doi:10.1016/j.cell.2018.10.001
104. Mills KHG. IL-17 and IL-17-producing cells in protection versus pathology. *Nat Rev Immunol*. 2023;23(1):38-54. doi:10.1038/s41577-022-00746-9
105. Tamassia N, Arruda-Silva F, Wright HL, et al. Human neutrophils activated via TLR8 promote Th17 polarization through IL-23. *J Leukoc Biol*. 2019;105(6):1155-1165. doi:10.1002/JLB.MA0818-308R
106. Keir HR, Chalmers JD. Neutrophil extracellular traps in chronic lung disease: implications for pathogenesis and therapy. *Eur Respir Rev*. 2022;31(163):210241. doi:10.1183/16000617.0241-2021
107. Croxatto D, Micheletti A, Montaldo E, et al. Group 3 innate lymphoid cells regulate neutrophil migration and function in human decidua. *Mucosal Immunol*. 2016;9(6):1372-1383. doi:10.1038/mi.2016.10
108. Pichler WJ, Hausmann O. Classification of Drug Hypersensitivity into Allergic, p-i, and Pseudo-Allergic Forms. *Int Arch Allergy Immunol*. 2016;171(3-4):166-179. doi:10.1159/000453265
109. Czarnowicki T, Gonzalez J, Shemer A, et al. Severe atopic dermatitis is characterized by selective expansion of circulating TH2/TC2 and TH22/TC22, but not TH17/TC17, cells within the skin-homing T-cell population. *J Allergy Clin Immunol*. 2015;136(1):104-115.e7. doi:10.1016/j.jaci.2015.01.020
110. Jones CP, Gregory LG, Causton B, Campbell GA, Lloyd CM. Activin a and TGF- $\beta$  promote T(H)9 cell-mediated pulmonary allergic pathology. *J Allergy Clin Immunol*. 2012;129(4):1000-10.e3. doi:10.1016/j.jaci.2011.12.965
111. Lu LF, Lind EF, Gondek DC, et al. Mast cells are essential intermediaries in regulatory T-cell tolerance. *Nature*. 2006;442(7106):997-1002. doi:10.1038/nature05010
112. Licona-Limón P, Henao-Mejia J, Temann AU, et al. Th9 cells drive host immunity against gastrointestinal Worm infection. *Immunity*. 2013;39(4):744-757. doi:10.1016/j.immuni.2013.07.020
113. Xiao X, Balasubramanian S, Liu W, et al. OX40 signaling favors the induction of T(H)9 cells and airway inflammation. *Nat Immunol*. 2012;13(10):981-990. doi:10.1038/ni.2390
114. Elyaman W, Bradshaw EM, Uyttenhove C, et al. IL-9 induces differentiation of TH17 cells and enhances function of FoxP3+ natural regulatory T cells. *Proc Natl Acad Sci U S A*. 2009;106(31):12885-12890. doi:10.1073/pnas.0812530106
115. Dugas B, Renaud JC, Pène J, et al. Interleukin-9 potentiates the interleukin-4-induced immunoglobulin (IgG, IgM and IgE) production by normal human B lymphocytes. *Eur J Immunol*. 1993;23(7):1687-1692. doi:10.1002/eji.1830230743
116. Matsuzawa S, Sakashita K, Kinoshita T, Ito S, Yamashita T, Koike K. IL-9 enhances the growth of human mast cell progenitors under stimulation with stem cell factor. *J Immunol*. 2003;170(7):3461-3467. doi:10.4049/jimmunol.170.7.3461
117. Nakagome K, Imamura M, Kawahata K, et al. High expression of IL-22 suppresses antigen-induced immune responses and eosinophilic airway inflammation via an IL-10-associated mechanism. *J Immunol*. 2011;187(10):5077-5089. doi:10.4049/jimmunol.1001560
118. Johnson JR, Nishioka M, Chakir J, et al. IL-22 contributes to TGF- $\beta$ 1-mediated epithelial-mesenchymal transition in asthmatic bronchial epithelial cells. *Respir Res*. 2013;14(1):118. doi:10.1186/1465-9921-14-118
119. Pennino D, Bhavsar PK, Effner R, et al. IL-22 suppresses IFN- $\gamma$ -mediated lung inflammation in asthmatic patients. *J Allergy Clin Immunol*. 2013;131(2):562-570. doi:10.1016/j.jaci.2012.09.036
120. Dudakov JA, Hanash AM, van den Brink MR. Interleukin-22: immunobiology and pathology. *Annu Rev Immunol*. 2015;33:747-785. doi:10.1146/annurev-immunol-032414-112123
121. Eguiluz-Gracia I, Tay TR, Hew M, et al. Recent developments and highlights in biomarkers in allergic diseases and asthma. *Allergy*. 2018;73(12):2290-2305. doi:10.1111/all.13628

122. Akdis CA. Does the epithelial barrier hypothesis explain the increase in allergy, autoimmunity and other chronic conditions? *Nat Rev Immunol*. 2021;21(11):739-751. doi:10.1038/s41577-021-00538-7
123. Schleimer RP, Berdnikovs S. Etiology of epithelial barrier dysfunction in patients with type 2 inflammatory diseases. *J Allergy Clin Immunol*. 2017;139(6):1752-1761. doi:10.1016/j.jaci.2017.04.010
124. Desai MS, Seekatz AM, Koropatkin NM, et al. A dietary fiber-deprived gut microbiota degrades the colonic mucus barrier and enhances pathogen susceptibility. *Cell*. 2016;167(5):1339-1353.e21. doi:10.1016/j.cell.2016.10.043
125. Parrish A, Boudaud M, Kuehn A, Ollert M, Desai MS. Intestinal mucus barrier: a missing piece of the puzzle in food allergy. *Trends Mol Med*. 2022;28(1):36-50. doi:10.1016/j.molmed.2021.10.004
126. Palmer CN, Irvine AD, Terron-Kwiatkowski A, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet*. 2006;38(4):441-446. doi:10.1038/ng1767
127. Irvine AD, McLean WH, Leung DY. Filaggrin mutations associated with skin and allergic diseases. *N Engl J Med*. 2011;365(14):1315-1327. doi:10.1056/NEJMra1011040
128. Nakamura M, Kamiya K, Furuhashi A, Ikeda K, Niyonsaba F. S100A7 Co-localization and up-regulation of Filaggrin in human Sinonasal epithelial cells. *Curr Med Sci*. 2021;41(5):863-868. doi:10.1007/s11596-021-2431-1
129. Soyka MB, Wawrzyniak P, Eiwegger T, et al. Defective epithelial barrier in chronic rhinosinusitis: the regulation of tight junctions by IFN- $\gamma$  and IL-4. *J Allergy Clin Immunol*. 2012;130(5):1087-1096.e10. doi:10.1016/j.jaci.2012.05.052
130. de Kleer IM, Kool M, de Bruijn MJ, et al. Perinatal activation of the Interleukin-33 pathway promotes type 2 immunity in the developing lung. *Immunity*. 2016;45(6):1285-1298. doi:10.1016/j.immuni.2016.10.031
131. Hiraishi Y, Yamaguchi S, Yoshizaki T, et al. IL-33, IL-25 and TSLP contribute to development of fungal-associated protease-induced innate-type airway inflammation. *Sci Rep*. 2018;8(1):18052. doi:10.1038/s41598-018-36440-x
132. Steelant B, Farré R, Wawrzyniak P, et al. Impaired barrier function in patients with house dust mite-induced allergic rhinitis is accompanied by decreased occludin and zonula occludens-1 expression. *J Allergy Clin Immunol*. 2016;137(4):1043-1053.e5. doi:10.1016/j.jaci.2015.10.050
133. Celebi Sozener Z, Ozdel Ozturk B, Cerci P, et al. Epithelial barrier hypothesis: effect of the external exposome on the microbiome and epithelial barriers in allergic disease. *Allergy*. 2022;77(5):1418-1449. doi:10.1111/all.15240
134. Moloudizargari M, Moradkhani F, Asghari N, et al. NLRP inflammasome as a key role player in the pathogenesis of environmental toxicants. *Life Sci*. 2019 Aug;15(231):116585. doi:10.1016/j.lfs.2019.116585
135. Doyle AD, Masuda MY, Pyon GC, et al. Detergent exposure induces epithelial barrier dysfunction and eosinophilic inflammation in the esophagus. *Allergy*. 2023;78(1):192-201. doi:10.1111/all.15457
136. Haahtela T, Holgate S, Pawankar R, et al. WAO special committee on climate change and biodiversity. The biodiversity hypothesis and allergic disease: world allergy organization position statement. *World Allergy Organ J*. 2013;6(1):3. doi:10.1186/1939-4551-6-3
137. Levy M, Kolodziejczyk AA, Thaiss CA, Elinav E. Dysbiosis and the immune system. *Nat Rev Immunol*. 2017;17(4):219-232. doi:10.1038/nri.2017.7
138. Wood LG. Asthma in the obese: a big and growing problem. *Am J Respir Crit Care Med*. 2017;195(1):4-5. doi:10.1164/rccm.201608-1582ED
139. Forno E, Han YY, Mullen J, Celedón JC. Overweight, obesity, and lung function in children and adults—a meta-analysis. *J Allergy Clin Immunol Pract*. 2018;6(2):570-581.e10. doi:10.1016/j.jaip.2017.07.010
140. Sharma V, Cowan DC. Obesity, inflammation, and severe asthma: an update. *Curr Allergy Asthma Rep*. 2021;21(12):46. doi:10.1007/s11882-021-01024-9
141. Sunadome H, Matsumoto H, Izuhara Y, et al. Correlation between eosinophil count, its genetic background and body mass index: the Nagahama study. *Allergol Int*. 2020;69(1):46-52. doi:10.1016/j.alit.2019.05.012
142. Zheng H, Wu D, Wu X, et al. Leptin promotes allergic airway inflammation through targeting the unfolded protein response pathway. *Sci Rep*. 2018;8(1):8905. doi:10.1038/s41598-018-27278-4
143. Russo S, Kwiatkowski M, Govorukhina N, Bischoff R, Melgert BN. Meta-inflammation and metabolic reprogramming of macrophages in diabetes and obesity: the importance of metabolites. *Front Immunol*. 2021;12(12):746151. doi:10.3389/fimmu.2021.746151
144. Michalovich D, Rodriguez-Perez N, Smolinska S, et al. Obesity and disease severity magnify disturbed microbiome-immune interactions in asthma patients. *Nat Commun*. 2019;10(1):5711. doi:10.1038/s41467-019-13751-9
145. Raybould HE. Gut microbiota, epithelial function and derangements in obesity. *J Physiol*. 2012;590(3):441-446. doi:10.1111/jphysiol.2011.222133
146. Cox AJ, West NP, Cripps AW. Obesity, inflammation, and the gut microbiota. *Lancet Diabetes Endocrinol*. 2015;3(3):207-215. doi:10.1016/S2213-8587(14)70134-2
147. Smolinska S, Jutel M, Cramer R, O'Mahony L. Histamine and gut mucosal immune regulation. *Allergy*. 2014;69(3):273-281. doi:10.1111/all.12330
148. Barcik W, Pugin B, Brescò MS, et al. Bacterial secretion of histamine within the gut influences immune responses within the lung. *Allergy*. 2019;74(5):899-909. doi:10.1111/all.13709
149. Jutel M, Watanabe T, Klunker S, et al. Histamine regulates T-cell and antibody responses by differential expression of H1 and H2 receptors. *Nature*. 2001;413(6854):420-425. doi:10.1038/35096564
150. Forde B, Yao L, Shaha R, Murphy S, Lunjani N, O'Mahony L. Immunomodulation by foods and microbes: unravelling the molecular tango. *Allergy*. 2022;77(12):3513-3526. doi:10.1111/all.15455
151. Kowalski ML, Asero R, Bavbek S, et al. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*. 2013;68(10):1219-1232. doi:10.1111/all.12260
152. Doña I, Barrionuevo E, Salas M, et al. NSAIDs-hypersensitivity often induces a blended reaction pattern involving multiple organs. *Sci Rep*. 2018;8(1):16710. doi:10.1038/s41598-018-34668-1
153. Doña I, Jurado-Escobar R, Perkins JR, et al. Eicosanoid mediator profiles in different phenotypes of nonsteroidal anti-inflammatory drug-induced urticaria. *Allergy*. 2019;74(6):1135-1144. doi:10.1111/all.13725
154. Kowalski ML, Agache I, Bavbek S, et al. Diagnosis and management of NSAID-exacerbated respiratory disease (N-ERD)—a EAACI position paper. *Allergy*. 2019;74(1):28-39. doi:10.1111/all.13599
155. Taniguchi M, Mitsui C, Hayashi H, et al. Aspirin-exacerbated respiratory disease (AERD): current understanding of AERD. *Allergol Int*. 2019;68(3):289-295. doi:10.1016/j.alit.2019.05.001
156. White AA, Stevenson DD. Aspirin-exacerbated respiratory disease. *N Engl J Med*. 2018;379(11):1060-1070. doi:10.1056/NEJMra1712125
157. Hybar H, Saki N, Maleknia M, Moghaddasi M, Bordbar A, Naghavi M. Aspirin exacerbated respiratory disease (AERD): molecular and cellular diagnostic & prognostic approaches. *Mol Biol Rep*. 2021;48(3):2703-2711. doi:10.1007/s11033-021-06240-0

158. Lyly A, Laidlaw TM, Lundberg M. Pathomechanisms of AERD-recent advances. *Front Allergy*. 2021;2:734733. doi:10.3389/falgy.2021.734733
159. Kohanski MA, Cohen NA, Barrett NA. Epithelial dysregulation in chronic rhinosinusitis with nasal polyposis (CRSwNP) and aspirin-exacerbated respiratory disease (AERD). *J Allergy Clin Immunol*. 2021;148(5):1161-1164. doi:10.1016/j.jaci.2021.07.034
160. Zhang T, Che D, Liu R, et al. Typical antimicrobials induce mast cell degranulation and anaphylactoid reactions via MRGPRX2 and its murine homologue MRGPRB2. *Eur J Immunol*. 2017;47(11):1949-1958. doi:10.1002/eji.201746951
161. Fujisawa D, Kashiwakura J, Kita H, et al. Expression of mas-related gene X2 on mast cells is upregulated in the skin of patients with severe chronic urticaria. *J Allergy Clin Immunol*. 2014;134(3):622-633.e9. doi:10.1016/j.jaci.2014.05.004
162. McNeil BD, Pundir P, Meeker S, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature*. 2015;519(7542):237-241. doi:10.1038/nature14022
163. da Silva EZ, Jamur MC, Oliver C. Mast cell function: a new vision of an old cell. *J Histochem Cytochem*. 2014;62(10):698-738. doi:10.1369/0022155414545334
164. Prussin C, Metcalfe DD. 4. IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol*. 2003;111(2 Suppl):S486-S494. doi:10.1067/mai.2003.120
165. Jutel M, Mosnaim GS, Bernstein JA, et al. The one health approach for allergic diseases and asthma. *Allergy*. 2023;78(7):1777-1793. doi:10.1111/all.15755

[Correction added on 3 November 2023, after first online publication: the sequence and citations of references # 55 to 165 were incorrect and have been renumbered throughout the article and references in this version.]

**How to cite this article:** Jutel M, Agache I, Zemelka-Wiacek M, et al. Nomenclature of allergic diseases and hypersensitivity reactions: Adapted to modern needs: An EAACI position paper. *Allergy*. 2023;78:2851-2874. doi:10.1111/all.15889